Asymmetric Total Synthesis of Phosphatidylinositol 3-Phosphate and 4-Phosphate Derivatives

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New asymmetric syntheses of phosphatidylinositol 3-phosphate (PtdIns(3)P) and phosphatidylinositol 4-phosphate (PtdIns(4)P) derivatives are described. Key intermediates were used to prepare diacylglyceryl moieties with dibutyryl, dioctanoyl, and dihexadecanoyl chains. In addition, a modified route provided PtdIns(3)P and PtdIns(4)P triesters with P-1-linked aminopropyl groups for preparation of affinity probes. The synthesis of the inosityl precursor employed a dibutyltin oxide-mediated p-methoxybenzyl (PMB) etherification to give either the 2-PMB- or the 3-PMBprotected glucopyranosides. The Ferrier rearrangement was used to convert suitably protected glucose derivatives to enantiomerically pure, differentially protected D-myo-inositol key intermediates. A versatile phosphoramidite reagent was employed to allow synthesis of PtdInsP_n derivatives with diacylglyceryl moieties of different chain lengths.

Introduction

Phosphatidylinositol polyphosphates (PtdInsP_ns) are key signaling molecules in cellular communication via protein kinases in endo- and exocytosis and in vesicular trafficking of proteins.^{1,2} The central molecule, L-aphosphatidyl-D-myo-inositol 4,5-bis(phosphate) (PtdIns- $(4,5)P_2$) serves four different cellular roles, each utilizing different portions of the total molecule. First, PtdIns-(4,5)P₂ is a substrate for phospholipase C (PLC), releasing the calcium-mobilizing second messenger Ins(1,4,5)P₃.³ Second, PtdIns(4,5)P₂ itself functions in the recruitment of proteins to membranes via pleckstrin homology (PH) domains.⁴ Third, PtdIns(4,5)P₂ may bind to and regulate cytoskeletal signaling proteins such as gelsolin and profilin⁵ or it may regulate the GTPase function of ARF or the phospholipase activity of PLD.^{6,7} Fourth, PtdIns-(4,5)P₂ can be converted by agonist-stimulated, receptormediated activation of phosphoinositide 3-kinase (PI $(3-K)^8$ to PtdIns $(3,4,5)P_3$, an effector molecule in a new intracellular signaling system.9 The PI 3-K pathway and the corresponding D-3 PtdInsP_n products¹⁰ have been linked to mechanisms of oncogene transformation, cytoskeletal rearrangements, membrane association of signaling proteins, and trafficking of proteins by coated vesicles. Characteristics of the three families of PI 3-Ks were recently reviewed.11

(10) Toker, A.; Cantley, L. C. Nature 1997, 387, 673-676.

The phosphoinositide monophosphates PtdIns(3)P and PtdIns(4)P and their protein targets have been less extensively examined for their roles in cell signaling. Studies have revealed that PtdIns(3)P, the product of the PI 3-K *VPS34*, is required for proper targeting of newly synthesized and endocytosed macromolecules in yeast.¹ PtdIns(3)P and other D-3 phosphoinositides bind to proteins that regulate vesicle coating, such as assembly proteins AP-2¹² and AP-3 (also known as AP-180),¹³ and proteins that mediate communication between the actin cytoskeleton and membrane signaling proteins.^{14,15} PtdIns(3)P is also a substrate for a 5-kinase that produces PtdIns(3,5)P₂ in mammalian and yeast cells.^{16–18}

PtdIns(4)P is a substrate for PI 5-kinases, which may operate on substrate bound to phosphoinositide transport proteins, for the biosynthesis of PtdIns(4,5)P₂.¹⁹ Two specific PtdIns(4)P 5-kinases are known that will also phosphorylate D-3 phosphoinositides to give bis- and tris-(phosphate) products.^{20,21} In addition, PtdIns(4)P shows affinity for dynamin¹⁸ and modest inhibition of a PtdIns-(3,4,5)P₃ 5-phosphatase.¹⁶

One constraint for the exploration of the biological roles of PtdIns(3)P and PtdIns(4)P has been limited access to synthetic materials with defined acyl chains; commer-

[†] These authors contributed equally to this research.

⁽¹⁾ Decamilli, P.; Emr, S. D.; McPherson, P. S.; Novick, P. Science **1996**, 271, 1533-1539.

Schekman, R.; Orci, L. Science 1996, 271, 1526–1533.
 Berridge, M. J. Mol. Cell. Endocrinol. 1994, 98, 119–124.

⁽⁴⁾ Saraste, M.; Hyvönen, M. Curr. Opin. Struct. Biol. 1995, 5, 403-408

⁽⁵⁾ Janmey, P. Chem. Biol. 1995, 2, 61-65.

⁽⁶⁾ Morris, A. J.; Engebrecht, J. A.; Frohman, M. A. *Trends Pharmacol. Sci.* **1996**, *17*, 182–185.

⁽⁷⁾ Harmond, S. M.; Jenco, J. M.; Nakashima, S.; Cadwallader, K.; Gu, Q.-M.; Cook, S.; Nozawa, Y.; Prestwich, G. D.; Frohman, M. A.; Morris, A. J. J. Biol. Chem. **1997**, *272*, 3860–3868.

⁽⁸⁾ Stephens, L.; Cooke, F. T.; Walters, R.; Jackson, T.; Volinia, S.; Gout, I.; Waterfield, M. D.; Hawkins, P. T. *Curr. Biol.* **1994**, *4*, 203-214.

⁽⁹⁾ Stephens, L. R.; Jackson, T. R.; Hawkins, P. T. *Biochim. Biophys.* Acta **1993**, *1179*, 27–75.

⁽¹¹⁾ Vanhaesebroeck, B.; Leevers, S.; Panayotou, G.; Waterfield, M. TIBS 1997, 22, 267-272.

⁽¹²⁾ Gaidarov, I.; Chen, Q.; Falck, J. R.; Reddy, K. K.; Keen, J. H. J. Biol. Chem. 1996, 271, 20922-20929.

⁽¹³⁾ Hao, W. H.; Tan, Z.; Prasad, K.; Reddy, K. K.; Chen, J.; Prestwich, G. D.; Falck, J. R.; Shears, S. B.; Lafer, E. M. J. Biol. Chem. 1997, 272, 6393-6398.

⁽¹⁴⁾ Blader, I. J.; Profit, A. A.; Jackson, T. R.; Greenwood, A. F.; Prestwich, G. D.; Theibert, A. B. *Mol. Biol. Cell* **1998**, in press.

⁽¹⁵⁾ Hammonds-Odie, L. P.; Jackson, T. R.; Profit, A. A.; Blader, I. J.; Turck, C.; Prestwich, G. D.; Theibert, A. B. J. Biol. Chem. 1996, 271, 18859-18868.

⁽¹⁶⁾ Whiteford, C. C.; Brearley, C. A.; Ulug, E. T. Biochem. J. 1997, *323*, 597–601.

⁽¹⁷⁾ Tolias, K.; Rameh, L.; Ishihara, H.; Shibasaki, Y.; Chen, J.; Prestwich, G. D.; Cantley, L.; Carpenter, C. J. Biol. Chem. 1998, 273, 18040-18046.

⁽¹⁸⁾ Dove, S.; Cooke, F.; Douglas, M.; Sayers, L.; Parker, P.; Michell, R. *Nature* **1997**, *390*, 187–192.

⁽¹⁹⁾ Cunningham, E.; Thomas, G. M. H.; Ball, A.; Hiles, I.; Cockcroft, S. *Curr. Biol.* **1995**, *5*, 775–783.

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cially available materials from biological sources of PtdIns have unavoidable heterogeneity in the glyceryl acyl chains. Indeed, biologists have frequently resorted to comparisons of binding or activation by symmetrical dioctanoyl or dipalmitoyl PtdInsP_n derivatives with naturally occurring PtdInsP₁s, such as the sn-1-O-stearoyl-2-O-arachidonyl PtdIns $(4,5)P_2$, which is most abundant in animal systems. To continue our efforts to prepare a complete "matched set" of diacylglyceryl derivatives of the PtdInsP_ns, which all derive their D-myo-inosityl chirality retrosynthetically from α -D-glucose,²² we describe herein the preparation of PtdIns(3)P and PtdIns-(4)P with short (dibutyryl), medium (dioctanoyl), and long (dipalmitoyl) chains. We also describe the preparation of the corresponding P-1-linked aminopropyl triesters^{23,24} and their use for the preparation of photoactivatable reagents that probe the water-bilayer interface for ligand-protein interactions.²²

Results and Discussion

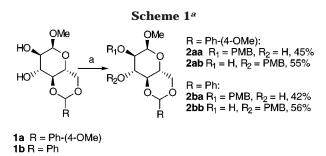
The strategy for the synthesis of enantiomerically pure PtdIns(3)P and PtdIns(4)P derivatives followed the route developed to synthesize soluble inositol polyphosphates $(InsP_n)$ and $PtdInsP_n$ congeners^{22–28} via the Ferrier rearrangement.^{22,29} For these syntheses, methyl D-aglucopyranoside was chosen as the optically pure starting material for the inositol moiety, and protecting groups are installed to establish the desired regiochemistry for the phosphate and hydroxyl groups. The key step involves the unique protection of the glucosyl C-2 OH or C-3 OH (corresponding to the C-3 OH or C-4 OH of D-*myo*-inositol) relative to the other hydroxyl groups; p-methoxybenzyl (PMB) was chosen for this purpose because it could be easily removed by DDQ without affecting other protecting groups.²² This protection strategy was accomplished using dibutyltin oxidemediated³⁰⁻³⁴ selective PMB protection of the 2-OH of methyl 4,6-O-(p-methoxybenzylidene)-D-α-glucopyranoside (1a), giving the 2-PMB ether 2aa in ca. 45% isolated yield (Scheme 1). Alternatively, the 3-PMB protected glycopyranoside **2ab** was obtained in 55% yield. The two isomers were readily separable by chromatography on silica gel. Use of either tetra-n-butylammonium bromide or iodide was necessary for optimal etherification and for

(20) Rameh, L.; Tolias, K.; Duckworth, B.; Cantley, L. Nature 1997, 390, 192-196.

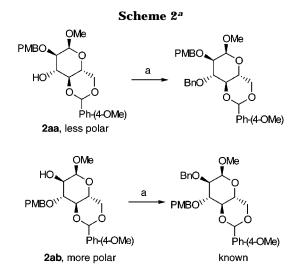
(21) Zhang, X. L.; Loijens, J. C.; Boronenkov, I. V.; Parker, G. J.; Norris, F. A.; Chen, J.; Thum, O.; Prestwich, G. D.; Majerus, P. W.; Anderson, R. A. J. Biol. Chem. 1997, 272, 17756-17761

(22) Prestwich, G. D. Acc. Chem. Res. 1996, 29, 503-513.

- (23) Gu, Q.-M.; Prestwich, G. D. J. Org. Chem. 1996, 61, 8642-8647.
- (24) Thum, O.; Chen, J.; Prestwich, G. D. Tetrahedron Lett. 1996, 37, 9017-9020.
- (25) Chen, J.; Profit, A. A.; Prestwich, G. D. J. Org. Chem. 1996, 61, 6305-6312
- (26) Chen, J.; Dormán, G.; Prestwich, G. D. J. Org. Chem. 1996, 61, 393-397. (27) Dormán, G.; Chen, J.; Prestwich, G. D. Tetrahedron Lett. 1995,
- 36, 8719-8722
- (28) Estevez, V. A.; Prestwich, G. D. J. Am. Chem. Soc. 1991, 113, 9885 - 9887
- (29) Ferrier, R.; Middleton, S. Chem. Rev. 1993, 93, 2779-2831.
- (30) Qin, H.; Grindley, T. B. J. Carbohydr. Chem. 1996, 15, 95-108.
- (31) Riley, A. M.; Jenkins, D. J.; Potter, B. V. L. J. Am. Chem. Soc. 1995, 117, 3300-3301.
- (32) Jenkins, D. J.; Potter, B. V. L. *Carbohydr. Res.* 1994, *265*, 145.
 (33) Jenkins, D. J.; Dubreuil, D.; Potter, B. V. L. *J. Chem. Soc.*, Perkin Trans. 1 1996, 1365-1372.
- (34) David, S.; Hanessian, S. Tetrahedron 1985, 41, 643-663.



^a Reaction conditions: (a) Bu₂SnO, *n*-Bu₄NBr, or *n*-Bu₄NI, toluene, reflux for 2 h; then PMBCl, reflux 2 h to overnight.



^a Reaction conditions: (a) NaH, BnBr, DMF, rt, 2h.

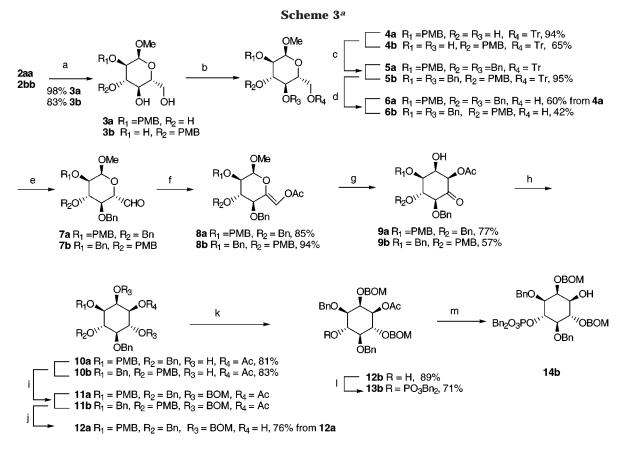
shorter reaction time. Although better regioselectivity has been reported using CH₃CN instead of toluene as solvent and by omission of the tetra-*n*-butylammonium salt,²⁸⁻³¹ lower yields and unacceptably long reaction times were experienced for a modest increase in regioselectivity. The 3-PMB ether 2ab was useful as the precursor for synthesis of Ins(1,4,5)P₃²⁷ and PtdIns(4,5)-P₂.^{23,25} Using the conditions described above, no 2,3-di-PMB ether was detected. Similar yields and regiospecificity were achieved from the commercially available methyl 4,6-O-benzylidene-D-α-glucopyranoside (1b).

To confirm the structures of the two isomers, a short chemical transformation was performed. Isomers 2aa and **2ab** were converted to benzyl ethers (Scheme 2). Comparison of the ¹H NMR and melting points with the known compounds²⁷ allowed identification of the less polar starting material as the 2-PMB ether 2aa and the more polar isomer as the 3-PMB ether 2ab.

As shown in Scheme 3, hydrolytic deprotection of the benzylidene acetal in 2aa and 2bb under mild acidic conditions unmasked the 4- and 6-hydroxyls (3a,b) in high yield without affecting the PMB group. The primary hydroxyl was selectively tritylated,^{28,35–37} the remaining secondary hydroxyls were benzylated, and mild acidic hydrolysis of the trityl group afforded alcohols 6a,b. Swern oxidation³⁸ gave aldehydes **7a**,**b**, which were

- many, 1994.
- (38) Mancuso, A. J.; Swern, P. Synthesis 1981, 165-185.

⁽³⁵⁾ Chaudhary, S. K.; Hernandez, O. Tetrahedron Lett. 1979, 95. (36) Greene, T. W.; Wuts, P. G. M. Protective Groups in Organic Synthesis, John Wiley & Sons: New York, 1991. (37) Kociénski, P. J. Protecting Groups, Thieme: Stuttgart, Ger-

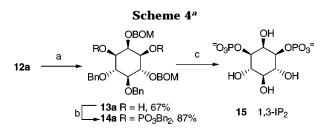


^{*a*} Reaction conditions: (a) 5% *p*-TsOH monohydrate (pH \sim 2), MeOH, rt, 1 h; (b) TrCl, DMAP (cat.), Et₃N, DMF, rt, overnight; (c) NaH, BnBr, DMF, rt, 2 h; (d) 5% *p*-TsOH monohydrate or sulfuric acid (pH \sim 2), MeOH, rt, 2 h; (e) oxalyl chloride, DMSO, CH₂Cl₂, -78 °C, then Et₃N, 1 h; (f) K₂CO₃, Ac₂O, CH₃CN, reflux, overnight; (g) Hg(OAc)₂, acetone–water (3:2), 45 min, then saturated NaCl solution, rt, overnight; (h) NaBH(OAc)₃, CH₃CN, HOAc, 45 min; (i) BOMCl, proton sponge, *n*-Bu₄NBr, rt, 8 h, then, 35 °C, overnight, then 55 °C, 10 h to overnight; (j) 0.35 M NaOH, MeOH, reflux, 2 h; (k) DDQ, CH₂Cl₂–H₂O (100:1, v/v), rt, 3 h; (l) (BnO)₂PNPr₂-*i*, 1*H*-tetrazole, rt, 30 min, then *m*-CPBA, -40 °C to room temperature, 60 min; (m) K₂CO₃, MeOH, 30 min.

directly acetylated to form the precursors **8a,b**, respectively. Ferrier rearrangement^{22,29} with Hg(OAc)₂ generated the inositol skeletons **9a,b**. Highly stereoselective reduction of the carbonyl group with triacetoxyborohydride³⁹ furnished the desired hydroxyl derivatives **10a,b**. Careful reaction²² of the diols **10a** and **10b** with BOMCl in the presence of proton sponge gave the bis-BOM compounds **11a,b** with no detectable acyl migration. DDQ removed the 4-PMB in **11b** to give a free OH, which was phosphorylated to triester **13b**; hydrolysis of the acetyl group then gave alcohol **14b**, which was ready for introduction of the phosphoglyceryl moiety.

We required the soluble $Ins(1,3)P_2$ for competitive displacement studies in the study of protein–PtdInsP_n interactions. This ligand was obtained (Scheme 4) by basic hydrolysis of the acetyl group of **11a**, removal of the PMB with DDQ, phosphorylation, and finally hydrogenolysis at 50 psi with Pd/C to give the *meso-myo*-inositol 1,3-bis(phosphate) **15**, in sodium salt form.⁴⁰

The differentially protected D-*myo*-inositol precursors to PtdIns(3)P and PtdIns(4)P, **12a** and **14b**, respectively, were employed for two sets of syntheses. In the first set (Scheme 5), the dipalmitoyl glyceryl affinity probes bearing the P-1-linked aminopropyl phosphotriester^{22–24} were prepared. Thus, coupling **12a** or **14b** with (cyano-



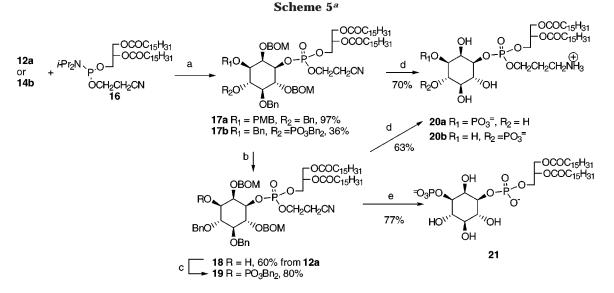
 a Reaction conditions: (a) DDQ, $CH_2Cl_2-H_2O$ (100:1, v/v), rt, 2 h; (b) (BnO)_2PNPr_2-*i*, 1*H*-tetrazole, rt, 30 min, then *m*-CPBA, –40 °C to room temperature, 60 min; (c) 10% Pd/C, NaHCO₃, *t*-BuOH–H_2O (6:1, v/v), H_2, 50 psi, rt, 5 h; then Chelex (Na^+ form) ion exchange.

ethyl)phosphoramidite **16** gave **17a**,**b**, respectively. Removal of PMB (from **17b**) with DDQ gave phosphate **18** in 60% yield in two steps. If ceric ammonium nitrate^{41,42} was used for oxidative deprotection of the PMB group, a slightly higher yield was obtained. Further phosphorylation furnished the fully protected phosphate **19** in 80% yield. The cyanoethyl group could be reduced to an aminopropyl group during hydrogenolysis of the benzyl protecting groups.²³ Thus, hydrogenation at 50 psi removed all the protecting groups of **17b** and **19** giving PtdIns(3)P and PtdIns(4)P triesters **20a**,**b**, respectively, as their sodium salts. Alternatively, initial removal of

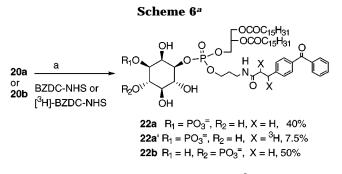
⁽³⁹⁾ Evans, D. A.; Chopman, K. T.; Carreira, E. M. J. Am. Chem. Soc. **1988**, *110*, 3560–3578.

⁽⁴⁰⁾ Billington, D. C.; Baker, R. J. Chem. Soc., Chem. Commun. 1987, 1011–1013.

⁽⁴¹⁾ Wang, Y.; Babirad, S. A.; Kishi, Y. J. Org. Chem. 1992, 57, 468.
(42) Johansson, R.; Samulsson, B. J. Chem. Soc., Perkin Trans. 1
1984, 2371–2374.



^{*a*} Reaction conditions: (a) 1*H*-tetrazole, rt, 30 min, then *m*CPBA, -40 °C to room temperature, 60 min; (b) DDQ, $CH_2Cl_2-H_2O$ (100:1, v/v), rt, 2 h; (c) (BnO)₂PNPr₂-*i*, 1*H*-tetrazole, rt, 30 min, then *m*-CPBA, -40 °C to room temperature, 60 min; (d) 10% Pd/C, NaHCO₃, *t*-BuOH-H₂O (6:1, v/v), H₂, 50 psi, rt, 5 h; (e) *i*-Pr₂NEt, MeOH, rt, overnight, then 10% Pd/C, NaHCO₃, *t*-BuOH-H₂O (6:1, v/v), H₂, 50 psi, rt, 5 h; (e) *i*-Pr₂NEt, MeOH, rt, overnight, then 10% Pd/C, NaHCO₃, *t*-BuOH-H₂O (6:1, v/v), H₂, 50 psi, rt, 5 h.



 $[^]a$ Reaction conditions: (a) BZDC–NHS or $[^3H]$ -BZDC–NHS, Et_3N, DMF, rt, overnight; then DEAE chromatography.

the cyanoethyl group of **19** with *i*-Pr₂NEt²³ followed by hydrogenation gave the desired PtdIns(3)P diester **21** in 77% yield.⁴³

The aminopropyl triesters were used in the preparation of photoactivatable probes for the proteins recognizing the inositol headgroup in the context of a lipid bilayer.²² Thus, reaction of the triester **20a** with unlabeled *p*benzoyldihydrocinnamoyl *N*-hydroxysuccinimide ester (BZDC–NHS) in DMF using Et₃N as base gave BZDC*triester*-PtdIns(3)P (**22a**) in ca. 50% yield.⁴⁴ When high specific activity [³H]BZDC–NHS was employed in a mixed 0.25 M TEAB buffer–DMF solvent system, the tritium-labeled **22a**' was obtained in 8% radiochemical yield (Scheme 6).

The long-chain diacyl derivatives of the phosphatidylinositol mono- and bis(polyphosphate)s are insoluble in water and form micelles and vesicles. However, the dioctanoyl analogues of PtdInsP₂ and PtdInsP₃ regioisomers are water soluble. It was unknown if the octanoyl derivative of PtdInsP regioisomers would be water soluble. To explore a variety of potential symmetrical diacylglyceryl derivatives from a common PtdInsP precursor, we prepared a versatile phosphoramidite (Scheme 7). Thus, glycerylphosphoramidite **23**⁴⁵ was prepared from reaction of benzyl N,N,N,N-tetraisopropylphosphoramidite with (S)-(+)-1,2-O-isopropylidene-sn-glycerol and showed chemical properties similar to those reported for similar phosphoramidites.⁴⁶ Reaction of inositol-protected 3-PMB intermediate 12 with the protected glycerylphosphoramidite 23 gave phosphate 24 in good yield. Removal of PMB (DDQ) and further phosphorylation gave bis-(phosphotriester) 26 in 60% yield. Hydrolysis of the acetal gave diol 27,47 which could be readily acylated to give a fully protected PtdIns(3)P precursor. As an example, we coupled 27 with octanoic acid and butyric acid in the presence of DCC and DMAP^{48,49} to give the protected products 28a,b, respectively, in high yields. Hydrogenation gave PtdIns(3)P derivatives 29a,b in 63% and 92% yields, respectively.

The ¹H NMR of **21**, the dipalmitoyl derivative of PtdIns(3)P, illustrates the curious solubility of the PtdInsP materials and the intrinsic difficulty in obtaining high-quality NMR spectroscopic data. In CDCl₃, the ¹H NMR resonances of the acylglyceryl moiety are reasonably well resolved, while only broad, poorly resolved resonances can be detected for the inositol phosphate headgroup. In contrast, in D_2O , the inositol phosphate headgroup becomes better-resolved, while the resonances for the fatty acyl chains and the glyceryl moiety become severely broadened. Furthermore, ³¹P NMR in D₂O showed one very broad resonance of poor diagnostic utility. Use of other solvents, e.g., DMSO- d_6 , makes recovery of precious materials for biological studies not feasible. Mixed solvents containing D₂O, CD₃OD, and CDCl₃ did not improve the situation. Changing cations to ammonium or to the acidic form also simply made spectra worse. In contrast, the dibutyryl and dioctanoyl

⁽⁴³⁾ Bruzik, K. S.; Kubiak, R. J. Tetrahedron Lett. 1995, 36, 2415–2418.

 ⁽⁴⁴⁾ Olszewski, J. D.; Dormán, G.; Elliott, J. T.; Hong, Y.; Ahern,
 D. G.; Prestwich, G. D. *Bioconjugate Chem.* 1995, *6*, 395–400.

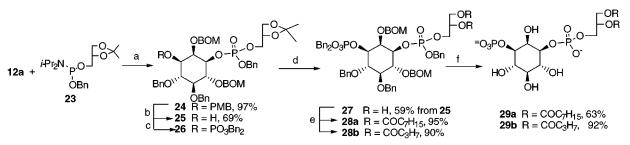
⁽⁴⁵⁾ Lemmen, P.; Buchweitz, K. M.; Stumpf, R. *Chem. Phys. Lipids* **1990**, *53*, 65–75.

⁽⁴⁶⁾ Beaucage, S. L.; Iyer, R. P. *Tetrahedron* **1993**, *49*, 10441–10488. (47) Heeb, N. V.; Nambiar, K. P. *Tetrahedron Lett.* **1993**, *34*, 6193–6196.

⁽⁴⁸⁾ Martin, S. F.; Wong, Y.-L.; Wagman, A. J. Org. Chem. 1994, 59, 4821–4831.

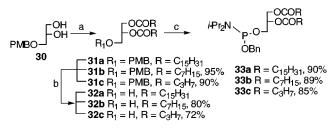
⁽⁴⁹⁾ Vodovozova, E. L.; Molotkovsky, J. G. *Tetrahedron Lett.* **1994**, *35*, 1933–1936.

Scheme 7^a



^{*a*} Reaction conditions: (a) 1*H*-tetrazole, rt, 30 min, then *m*CPBA, -40 °C to room temperature, 60 min; (b) DDQ, CH₂Cl₂-H₂O (100:1, v/v), rt, 2 h; (c) (BnO)₂PNPr₂-*i*, 1*H*-tetrazole, rt, 30 min, then *m*-CPBA, -40 °C to room temperature, 60 min; (d) 5% *p*-TsOH monohydrate (pH ~ 2), MeOH, rt, 2 h; (e) fatty acid, DMAP (cat.), DCC, CH₂Cl₂, rt, overnight; (f) 10% Pd/C, NaHCO₃, *t*-BuOH-H₂O (6:1, v/v), H₂, 50 psi, rt, 5 h; then Chelex (Na⁺ form) ion exchange.

Scheme 8^a

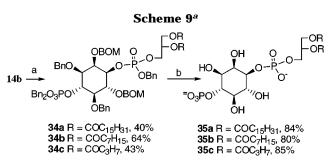


^{*a*} Reaction conditions: (a) Fatty acid, DMAP (cat.), DCC, CH_2CI_2 , rt, overnight; (b) DDQ, $CH_2CI_2-H_2O$ (100:1, v/v), rt, 2 h; (c) (BnO)P(Cl)(NPr₂-*i*), *i*-Pr₂NEt, CH_2CI_2 , 0 °C, 2 h.

derivatives were water soluble and gave high-resolution ¹H and ³¹P NMR spectra samples with a concentration of 10 mg/mL. The narrow ³¹P lines (0.028-0.075 ppm at half-height) for both dibutyryl and dioctanoyl PtdIns-(3)P and PtdIns(4)P suggest monomeric and not micellar forms at this concentration.

In the preparation of the series of PtdIns(4)P diesters, an alternative strategy of using different phosphoramidites with preattached long, medium, and short diacyl chains was followed (Scheme 8). Following reported procedures,²³⁻²⁵ the 3-O-PMB-sn-glycerol **30** was esterified with various fatty acids using DCC/DMAP to give compounds 31a-c in high yields. Oxidative removal of the PMB groups with DDQ gave the corresponding alcohols **32a**–**c** in good yields with no evidence of acyl migration. Finally, coupling these alcohols with benzyl N.N-isopropylchlorophosphoamidite gave reagents **33a**-c in high yields. Subsequent coupling of 4-phosphorylated inosityl precursor 14b with 33a, 33b, or 33c followed by mild oxidation gave the three protected PtdIns(4)P precursors **34a**-**c** in moderate to good yields (Scheme 9). Hydrogenolysis provided PtdIns(4)P derivatives with dipalmitoyl (35a), dioctanoyl (35b), and dibutyryl (35c) acyl chains. Both diester and triester with dipalmitoyl glyceryl groups showed poor solubility in water and good solubility in methanol-chloroform-water mixtures, while the dibutyryl and dioctanoyl derivatives were water soluble.

Recently, a variety of synthetic approaches to PtdInsP_ns have been summarized.^{22,50} Several recent approaches to PtdIns(3)P include preparation from D-1-*Otert*-butyldiphenylsilyl-*myo*-inositol⁴³ or from enzymatically resolved 1,2:5,6-di-*O*-cyclohexylidene-*myo*-inositol.⁵¹ The



^{*a*} Reaction conditions: (a) 1*H*-tetrazole, rt, 30 min, then *m*-CPBA, -40 °C to room temperature, 60 min; (b) 10% Pd/C, NaHCO₃, *t*-BuOH–H₂O (6:1, v/v), H₂, 50 psi, rt, 5 h; then Chelex (Na⁺ form) ion exchange.

soluble Ins(1,3)P₂ headgroup was synthesized from racemic myo-inositol derivatives.40,52 The synthesis reported herein now allows preparation of diester reagents or triester affinity probes²² for further examination of the roles of these phosphoinositide monophosphates in cell signaling. Applications of the [³H]BZDC-PtdIns(3)P and -PtdIns(4)P photoaffinity labels for the identification of new target proteins will be presented in due course. Finally, with these syntheses and the synthesis of both PtdIns(5)P and PtdIns(3,5)P₂,⁵³ biological researchers may now select from a complete "matched set" of dibutryl, dioctanoyl, or dipalmitoyl PtdIns P_n derivatives with either complete water solubility or complete bilayer localization to determine the preferred number and regiochemistry of phosphates required for optimal biological activity.

Experimental Section

NMR spectra (¹H, ¹³C, and ³¹P) were recorded in CDCl₃ or D₂O on QE-300, AC-250, or AC-200 NMR spectrometers, and shifts are reported relative to δ (TMS) = 0 ppm. When necessary, solvents and reagents were dried using standard procedures. Elemental analysis was performed by M-H-W Laboratories (Phoenix, AZ), and the mass spectra (low solution and high resolution) were measured at The University of California at Riverside (Riverside, CA) or at The University of Utah (Salt Lake City, UT) using chemical ionization (CI, with NH₃ giving MNH₄⁺ peaks), EI, or FAB (BNA as matrix giving MNa⁺ peaks). Chemicals from Aldrich Chemical Co. (Milwaukee, WI) and Sigma Chemical Co. (St. Louis, MO) were used without further purification. TLC (SiO₂) plates for inositol precursors were developed in ethyl acetate (EtOAc)

⁽⁵⁰⁾ Potter, B. V. L.; Lampe, D. Angew. Chem., Int. Ed. Engl. 1995, 34, 1933–1972.

⁽⁵¹⁾ Wang, D.-S.; Chen, C.-S. J. Org. Chem. 1996, 61, 5905-5910.

⁽⁵²⁾ Maryanoff, B. E.; Reitz, A. B.; Tutwiler, G. F.; Benkovic, S. J.; Benkovic, P. A.; Pilkis, S. J. J. Am. Chem. Soc. **1984**, 106, 7851.

⁽⁵³⁾ Peng, J.; Prestwich, G. D. *Tetrahedron Lett.* **1998**, *39*, 3965–3968.

and hexane mixtures and visualized by dipping in 5% phosphomolybdic acid in absolute ethanol followed by heating; for acylglycerol derivatives, compounds were visualized by dipping in anisaldehyde stains (ratio *p*-anisaldehyde:acetic acid:sulfuric acid:ethanol 2.5:1:3.5:93, v/v/v/v) followed by heating.

Methyl 2-O-(*p*-Methoxybenzyl)-4,6-O-(*p*-methoxybenzylidene)- α -D-glucopyranoside (2aa). A mixture of methyl 4,6-O-(*p*-methoxybenzylidene)- α -D-glucopyranoside (1a, 5.21 g, 16.7 mmol), Bu₂SnO (4.2 g, 16.9 mmol), and Bu₄NI (0.6 g, 1.63 mmol) in 100 mL of toluene was refluxed for 2 h, then *p*-methoxybenzyl chloride (PMBCl, 2.88 g, 18.3 mmol) was added, and the mixture was stirred under reflux for 2 h, at which point TLC showed no starting material. The mixture was cooled, diluted with EtOAc, washed (H₂O, brine), dried (MgSO₄), and concentrated in vacuo to a brown oil. Chromatography on SiO₂ (50% EtOAc-hexane) gave two products at $R_{f^{\sim}}$ 0.7. The first eluting product was the 2-PMB ether (2aa, 3.30 g, yield 45%), and the second product was 3-PMB ether (2ab, 3.8 g, 55% yield); the latter was less soluble in EtOAchexane. No bis-PMB product was detected.

Data for compound **2aa**: mp 108–110 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.36, 7.34, 7.26, 7.23 (4s, 4H, aromatic), 6.84– 6.81 (m, 4H, aromatic), 5.41 (s, 1H, CH), 4.61 (q, J = 11.7 Hz, 2H), 4.50 (d, J = 3.6 Hz, 1H), 4.15 (dd, J = 4.5, 4.2 Hz, 1H), 4.05 (dt, J = 9.2, 1.8 Hz, 1H), 3.74, 3.37 (2s, 6H, OMe), 3.43– 3.35 (m, 2H), 3.31 (s, 3H, OMe) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 129.9, 129.8, 129.6, 127.6, 113.9, 113.6, 101.8, 98.7, 81.2, 79.1, 77.2, 73.0, 70.2, 68.9, 62.0, 55.4, 55.3 ppm. Anal. Calcd for C₂₃H₂₈O₈: C, 63.88; H, 6.53. Found: C, 63.64; H, 6.77.

Methyl 3-*O*-(*p*-methoxybenzyl)-4,6-*O*-(*p*-methoxybenzylidene)-α-**D**-glucopyranoside (2ab) was isolated from the chromatography above: mp 163–165 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.40–7.20 (4s, 4H, aromatic), 6.84–6.81 (m, 4H, aromatic), 5.51 (s, 1H, CH), 4.81 (q, J = 11.7 Hz, 2H), 4.78 (d, J = 3.6 Hz, 1H), 4.30 (q, J = 3.5 Hz, 1H), 3.74, 3.37 (2s, 6H, OMe), 3.43–3.35 (m, 2H), 3.31 (s, 3H, OMe) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 129.7, 129.0, 128.3, 126.0, 113.8, 101.3, 98.7, 82.0, 78.4, 77.2, 73.0, 70.2, 69.0, 62.6, 55.4, 55.3 ppm. Anal. Calcd for C₂₃H₂₈O₈: C, 63.88; H, 6.53. Found: C, 63.53; H, 6.45.

Methyl 2-*O*-(*p*-Methoxybenzyl)-4,6-*O*-benzylidene- α -D-glucopyranoside (2ba). A mixture of methyl 4,6-*O*-benzylidene- α -D-glucopyranoside (1b, 25 g, 88.6 mmol), and Bu₂SnO (22 g, 88.6 mmol), Bu₄NBr (2.86 g, 8.86 mmol) in 300 mL of toluene was refluxed for 3 h, then PMBCl (16.7 g, 106.3 mmol) was added, and the mixture was stirred under reflux overnight. After workup as above and SiO₂ chromatography with 50% EtOAc-hexane, two products were isolated. The first eluting product ($R_f \sim 0.7$) was identified as methyl 2-*O*-(*p*-methoxybenzyl)-4,6-*O*-benzylidene- α -D-glucopyranoside (2-PMB ether **2ba**) (15 g, yield 42%). The second eluting compound was methyl 3-*O*-(*p*-methoxybenzyl)-4,6-*O*-benzylidene- α -D-glucopyranoside (3-PMB ether **2bb**) (20 g, 56% yield), which was less soluble in EtOAc-hexane mixtures and easier to recrystallize.

Data for compound **2ba**: ¹H NMR (300 MHz, CDCl₃) δ 7.60– 7.20 (m, 7H, aromatic), 6.90, 6.86 (2s, 2H, PMB), 5.50 (s, 1H, CH), 4.63 (q, J = 11.8 Hz, 2H), 4.55 (d, J = 3.6 Hz, 1H), 4.25 (dd, J = 4.5, 4.2 Hz, 1H), 4.10 (t, J = 9.2 Hz, 1H), 3.76 (s, 3H, OMe), 3.84–3.75 (m, 1H), 3.70 (q, J = 10.1 Hz, 1H), 3.43– 3.35 (m, 2H), 3.31 (s, 3H, OMe) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 130.0, 129.8, 129.2, 128.3, 126.4, 114.0, 101.9, 98.7, 81.3, 79.2, 73.0, 70.2, 69.0, 62.1, 55.4, 55.3 ppm. Anal. Calcd for C₂₂H₂₆O₇: C, 65.66; H, 6.51. Found: C, 65.40; H, 6.55.

Data for methyl 3-*O*-(*p*-methoxybenzyl)-4,6-*O*-benzylidene-α-D-glucopyranoside (2bb): ¹H NMR (300 MHz, CDCl₃) δ 7.60–7.30 (m, 7H, aromatic), 6.86, 6.82 (2s, 2H, PMB), 5.56 (s, 1H, CH), 4.81 (q, J = 11.7 Hz, 2H), 4.78 (d, J = 3.6 Hz, 1H), 4.30 (q, J = 3.5 Hz, 1H), 3.78 (s, 3H, OMe), 3.90–3.60 (m, 2H), 3.31 (s, 3H, OMe), 1.60 (br s, 1H, OH) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 129.7, 129.0, 128.3, 126.0, 113.8, 101.3, 98.7, 82.0, 78.4, 77.2, 73.0, 70.2, 69.0, 62.6, 55.4, 55.3 ppm. Anal. Calcd for C₂₂H₂₆O₇: C, 65.66; H, 6.51. Found: C, 65.46; H, 6.52.

Methyl 2-O-(p-Methoxybenzyl)-α-D-glucopyranoside (3a). A mixture of methyl 2-O-(p-methoxybenzyl)-4,6-O-(pmethoxybenzylidene)-a-D-glucopyranoside (2aa, 4 g, 9.25 mmol) and p-TsOH monohydrate (400 mg) in 100 mL of MeOH was stirred for 1 h. TLC ($R_f \sim 0.5$), ether) showed no starting material. Then, 10 mL of 10% NaHCO3 solution was added and the mixture was evaporated to dryness. The product was extracted five times from the solid material with EtOAc and then recrystallized from EtOAc-hexane to give 2.85 g of pure methyl 2-O-(p-methoxybenzyl)-α-D-glucopyranoside 3a (98%): ¹H NMR (300 MHz, CDCl₃) & 7.22, 7.19, 6.82, 6.79 (4s, 4H, aromatic), 4.56 (AB type, $J_{AB} = 11.4$ Hz, 2H, CH₂O), 4.48 (s, 1H), 3.85-3.70 (m, 2H), 3.72 (s, 3H, OMe), 3.61 (s, 1H), 3.49 (s, 2H), 3.37 (s, 2H), 3.25 (s, 3H, OMe), 2.52 (s, 1H), 2.01 (s, 1H); $^{13}\mathrm{C}$ NMR (75 MHz, CDCl₃) δ 129.9, 129.7, 113.9, 97.9, 79.0, 77.2, 72.8, 72.7, 55.2 ppm. Anal. Calcd for C₁₅H₂₂O₇: C, 57.31; H, 7.06. Found: C, 57.47; H, 6.79.

Methyl 3-O-(p-Methoxybenzyl)-α-D-glucopyranoside (3b). A mixture of methyl 3-O-(p-methoxybenzyl)-4,6-Obenzylidene- α -D-glucopyranoside (**2bb**, 11.0 g, 27.3 mmol) and p-TsOH monohydrate (930 mg) in 250 mL of MeOH was stirred for 2 h. Then, 2 g of NaHCO₃ was added, the NaHCO₃ solid was removed by filtration, and the filtrate was evaporated to dryness. The crude product was extracted from the residue with five portions of EtOAc, concentrated, and then purified on SiO₂ (75% EtOAc-hexane) to give 7.5 g of pure methyl 3-O-(*p*-methoxybenzyl)- α -D-glucopyranoside **3b** (83%) with $R_f \sim$ 0.21 (20% EtOAc-hexane): ¹H NMR (200 MHz, CDCl₃) δ 7.31, 7.28, 6.89, 6.85 (4s, 4H, PMB), 4.78 ($J_{AB} = 11.2$ Hz, 2H, CH₂O), 4.72 (s, 1H), 3.78 (m, 5H), 3.57-3.54 (m, 4H), 3.40 (s, 3H, OMe), 3.02 (s, 1H), 2.45 (s, 1H); 13 C NMR (50 MHz, CDCl₃) δ 159.1, 129.5, 113.8, 106.5, 104.5, 99.4, 82.0, 74.4, 72.4, 70.9, 69.7, 61.7, 55.1 ppm. MS(EI⁺): m/e 314 (M⁺), 150, 137, 121. HRMS (EI⁺): calcd for C₁₅H₂₂O₇ (M⁺), m/e 314.1366; found, m/e 314.1349.

Methyl 2-O-(p-Methoxybenzyl)-6-O-trityl-α-D-glucopy**ranoside (4a).** A mixture of methyl 2-O-(p-methoxybenzyl)α-D-glucopyranoside (3a, 2.85 g, 9.10 mmol), trityl chloride (2.79 g, 1.0 mmol), DMAP (40 mg), and triethylamine (3 mL) in 100 mL of dry DMF was stirred overnight. TLC ($R_f \sim 0.3$, 50% EtOAc-CH₂Cl₂) showed essentially complete conversion. The mixture was then poured into water, extracted with EtOAc, concentrated, and purified (SiO₂) to give 4.79 g (94%) of methyl 2-O-(p-methoxybenzyl)-6-O-trityl-α-D-glucopyranoside (4a): ¹H NMR (300 MHz, CDCl₃) & 7.45-7.10 (m, 17H, aromatic), 6.79 (d, J = 9.6 Hz, 2H, PMB), 4.61-4.51 (m, 3H), 3.80 (t, J = 0.6 Hz, 1H), 3.73 (s, 3H, OMe), 3.64-3.59 (m, 2H), 3.43-3.20 (m, 3H), 3.31 (s, 3H, OMe), 3.00 (s, br, 1H, OH), 1.93 (s, 1H, OH) ppm; ¹³C NMR (75 MHz, CDCl₃) δ: 143.8, 130.1, 129.6, 128.6, 128.0, 127.8, 127.0, 113.9, 97.6, 86.8, 78.8, 77.3, 73.1, 72.6, 71.7, 69.7, 64.0, 55.2, 55.1 ppm. MS (EI⁺): m/e 556 (M⁺), 313, 243, 165, 121, 91. HRMS (EI⁺): calcd for C₃₄H₃₆O₇ (M⁺), m/e 556.2461; found, m/e 556.2467.

Methyl 3-O-(p-Methoxybenzyl)-6-O-trityl-a-D-glucopyranoside (4b). A mixture of methyl 3-O-(p-methoxybenzyl)α-D-glucopyranoside (3b, 7.1 g, 22.4 mmol), trityl chloride (6.3 g, 22.5 mmol), DMAP (317 mg), and triethylamine (4.77 mL) in 150 mL of dry DMF was stirred overnight. TLC showed essentially complete conversion. The mixture was then poured into water, extracted with EtOAc, washed (brine, water), dried (MgSO₄), and purified (SiO₂) to give 8.2 g (65%) of methyl 3-O-(*p*-methoxybenzyl)-6-*O*-trityl-α-D-glucopyranoside (**4b**): ¹H NMR (200 MHz, CDCl₃) δ 7.52–7.21 (m, 17H, Ph + PMB), 6.88 (d, 2H, J = 8.0 Hz, PMB), 4.91–4.72 (m, 3H), 3.79 (s, 3H, OMe), 3.76-3.57 (m, 4H), 3.47 (s, 3H, OMe), 3.41-3.37 (m, 2H), 2.46 (s, 1H, OH), 2.33 (d, 1H, J = 6.0 Hz, OH); ¹³C NMR (50 MHz, CDCl₃) δ 159.5, 143.6, 130.5, 129.5, 128.5, 127.7, 126.9, 113.8, 105.5, 99.1, 86.7, 82.2, 74.5, 72.3, 71.3, 70.0 63.7, 55.1, 54.9 ppm. MS (EI⁺): m/e 555 (M - H⁺), 313, 243, 165, 137, 121. ĤRMS (EI⁺): calcd for C₃₄H₃₆O₇ (M⁺), m/e 556.2461; found, m/e 556.2426.

Methyl 3-*O*-(*p*-Methoxybenzyl)-2,4-*O*-dibenzyl-6-*O*-trityl- α -D-glucopyranoside (5b). A mixture of methyl 3-*O*-(*p*methoxybenzyl)-6-*O*-trityl- α -D-glucopyranoside (4b, 8.0 g, 14.3 mmol) and NaH (1.5 g, ~60% oil, 37.5 mmol), and benzyl bromide (5.76 g, 33.6 mmol) in 150 mL of dry DMF was stirred for 2 h at room temperature. The reaction was quenched with methanol, poured into 150 mL of water, extracted (3 volumes of CH₂Cl₂), and purified on SiO₂ (33% EtOAc-hexane) to give 10 g of pure **5b** (95%), $R_f \sim 0.49$ (20% EtOAc-hexane): ¹H NMR (200 MHz, CDCl₃) δ 7.51–7.21 (m, 27H, Ph + PMB), 6.87 (d, J = 8.8 Hz, 2H, PMB), 4.93–4.71 (m, 6H), 4.31 (d, J = 10.6 Hz, 1H), 3.98 (t, J = 9.2 Hz, 1H), 3.86–3.81 (m, 4H), 3.69–3.59 (m, 2H), 3.55–3.49 (m, 1H), 3.47 (s, 3H, OMe), 3.19 (dd, J = 4.6, 10.0 Hz, 2H); ¹³C NMR (50 MHz, CDCl₃) δ : 159.0, 143.7, 138.2, 137.5, 130.7, 129.6, 128.6–127.3 (m), 126.7, 113.6, 108.5, 97.7, 86.1, 81.6, 80.0, 77.9, 75.5, 74.8, 73.2, 70.0, 62.4, 55.1, 54.8 ppm. MS (EI⁺): m/e 735 (M – H⁺), 645, 615, 493, 461, 243, 121, 91. HRMS (EI⁺): calcd for C₄₈H₄₈O₇, m/e 736.3400; found, m/e 736.3364.

Methyl 2-O-(p-Methoxybenzyl)-3,4-O-dibenzyl-a-D-glucopyranoside (6a). A mixture of methyl 2-O-(p-methoxybenzyl)-6-O-trityl-α-D-glucopyranoside (4a, 4.79 g, 8.6 mmol) and NaH (1.03 g, ~60% oil, 25.8 mmol) and benzyl bromide (4.4 g, 25.8 mmol) in 100 mL of dry DMF was stirred for 2 h. TLC ($R_f \sim 0.9$, 50% EtOAc-hexane) showed nearly complete conversion. Then the mixture was poured into water and extracted with EtOAc. The crude product 5a was dissolved in 100 mL of MeOH containing p-TsOH monohydrate (40 mg, to give pH \sim 2), and the mixture was stirred for 1 h. TLC (R_f ~ 0.45 , 50% EtOAc-hexane) showed no starting material. Then, 10 mL of 10% NaHCO3 solution was added and the mixture was evaporated to dryness. The product was extracted from the solid material with five portions of EtOAc, concentrated, and then purified on SiO₂ using 50% EtOAchexane as eluent to give 2.53 g of pure methyl 2-O-(pmethoxybenzyl)-4,5-O-dibenzyl-α-D-glucopyranoside (6a) (60% in two steps): ¹H NMR (300 MHz, CDCl₃) δ 7.40–7.10 (m, 12H, aromatic), 6.80 (d, J = 8.4 Hz, 2H, PMB), 4.93 (d, J = 11.1 Hz, 1H), 4.80 (AB type, J = 10.8 Hz, 1H), 4.68 (d, J = 12 Hz, 1H), 4.56 (t, J = 13.2, Hz, 2H), 4.46 (d, J = 3 Hz, 1H), 3.94 (t, J = 9.3 Hz, 1H), 3.75 (3H, OMe), 3.70–3.55 (m, 4H), 3.49– 3.40 (m, 2H), 3.20 (s, 3H, OMe), 1.65 (s, 1H, OH) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 138.7, 138.1, 130.2, 129.7, 128.4-127.3 (m), 113.8, 98.2, 81.9, 79.6, 77.4, 77.2, 75.7, 75.0, 73.0, 70.6, 61.8, 55.2, 55.1 ppm. MS (CI⁺, NH₃): m/e 512 (MNH₄⁺), 493, 461, 403, 373, 121, 108, 91. HRMS (CI+, NH₃): calcd for C₂₉H₃₈NO₇ (MNH₄⁺), *m/e* 512.2648; found, *m/e* 512.2632.

Methyl 3-O-(p-Methoxybenzyl)-2,4-O-dibenzyl-α-D-glucopyranoside (6b). Compound 5b in 100 mL of MeOH/ acetone containing 5% sulfuric acid (pH \sim 2) was stirred for 1 h. Then the reaction was quenched by 10% NaHCO₃ solution, the mixture was evaporated to dryness, and the residue was extracted with three volumes of EtOAc and the combined organic phases were washed with 10% NaHCO3. Purification on SiO₂ (25% EtOAc-hexane) gave 2.8 g of glucopyranoside **6b** (yield 42%), $R_f \sim 0.16$ (33% EtOAc-hexane): ¹H NMR (200 MHz, CDCl₃) δ 7.38–7.27 (m, 12H, Bn + PMB), 6.86 (d, J = 8.6 Hz, 2H, PMB), 4.95–4.57 (m, 7H), 4.01 (t, J=9.1 Hz, 1H), 3.80 (s, 3H, OMe), 3.76-3.63 (m, 3H), 3.56-3.47 (m, 2H), 3.37 (s, 3H, OMe), 1.80 (s, 1H, OH); 13 C NMR (50 MHz, CDCl₃) δ 159.0, 137.9, 131.0, 129.5, 128.3-127.7 (m), 113.6, 98.0, 81.5, 79.9, 76.2, 75.3, 74.9, 73.2, 70.1, 61.6, 55.1, 55.0 ppm. MS (EI⁺): m/e 493 (M – H⁺), 403, 373, 253, 175, 121, 91. HRMS (EI⁺): calcd for $C_{29}H_{34}O_7$ (M⁺), *m/e* 494.2305; found, *m/e* 494.2299

Methyl (*Z*)-3,4-*O*-Dibenzyl-2-*O*-(*p*-methoxybenzyl)-6-*O*acetyl- α -D-glucos-5-enopyranoside (8a). A solution of (COCl)₂ (1.05 mL, 1.94 mmol) in 100 mL of CH₂Cl₂ was cooled to -78 °C, dry DMSO (1.71 mL, 3.87 mmol) was added dropwise, and the mixture was stirred 10 min at -78 °C. Next, a solution of **6a** (2.40 g, 4.86 mmol) in 5 mL of CH₂Cl₂ was added over 10 min. The cloudy solution was stirred for 30 min, 5.4 mL of Et₃N was added to produce a clear solution, and the mixture was warmed to room temperature. The usual workup gave oil **7a** that was used directly in the next step. Thus, oil **7a** was dissolved in 75 mL of dry CH₃CN, anhydrous K₂CO₃ (4.13 g) was added, the mixture was stirred for 10 min, acetic anhydride (2.7 mL) was added, and the mixture was refluxed overnight under nitrogen. The volume was reduced to onethird, and the residue was diluted with water and extracted with ether. The usual workup gave 2.2 g of enol acetate **8a** (85% in two steps), $R_f \sim 0.6$, 50% EtOAc–hexane: ¹H NMR (250 MHz, CDCl₃) δ 7.4–7.2 (m, 12H), 6.80, 6.76 (2s, 2H, PMB), 4.90–4.60 (m, 8H), 4.00–3.90 (m, 2H), 3.80 (s, 3H, OMe), 3.60–3.50 (m, 1H), 3.35 (s, 3H), 2.20 (s, 3H) ppm; ¹³C NMR (63 MHz, CDCl₃) δ 167.3, 159.2, 153.3, 138.0, 135.1, 130.8, 129.8, 128.5, 128.1, 123.0, 113.9, 113.8, 99.8, 81.1, 79.1, 75.4, 74.2, 73.7, 60.4, 56.2, 55.3 ppm. MS (CI⁺, NH₃): *m/e* 552 (MNH₄⁺), 432, 383, 325, 211, 137, 121, 108. HRMS (CI⁺): calcd for C₃₁H₃₈NO₈ (MNH₄⁺), *m/e* 552.2597; found, *m/e* 552.2597.

Methyl (Z)-2,4-O-Dibenzyl-3-O-(p-methoxybenzyl)-6-Oacetyl-α-D-glucos-5-enopyranoside (8b). A solution of (COCl)₂ (1.29 mL, 14.78 mmol) in 50 mL of CH₂Cl₂ was cooled to -78 °C, dry DMSO (2.1 mL, 29.6 mmol) was added dropwise, the mixture was stirred at -78 °C for 10 min, and a solution of alcohol 6b (2.8 g, 5.6 mmol) in 5 mL of CH₂Cl₂ was added over 10 min. The milky solution was stirred for 30 min, 6.6 mL of Et₃N was added to give a clear solution, and the mixture was warmed to room temperature and washed $(2 \times 0.5 \text{ M KHSO}_4, \text{ water})$ to give crude aldehyde **7b.** The crude product was dissolved in 50 mL of dry CH₃CN, and anhydrous K₂CO₃ (5.25 g) was added. After the mixture was stirred for 10 min, acetic anhydride (3.0 mL) was added and the mixture was refluxed overnight under nitrogen. The volume was reduced by rotary evaporation, and the residue was diluted with water, extracted with ether, and purified on SiO₂ (33% EtOAc-hexane) to give 2.6 g of enol acetate 8b (94% in two steps): ¹H NMR (200 MHz, CDCl₃) δ 7.39–7.17 (m, 12H, Bn + PMB), 6.86 (d, 2H, J = 8.7 Hz, PMB), 4.95–4.64 (m, 7H), 3.98-3.95 (m, 1H), 3.89-3.79 (m, 4H), 3.60-3.55 (m, 2H), 3.47 (s, 3H, OMe), 2.16 (s, 3H, Ac) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 185.9, 159.0, 137.4, 134.8, 129.5–127.7 (m), 122.8, 113.7, 113.6, 108.5, 107.5, 105.5, 99.6, 80.6, 78.8, 75.2, 74.3, 73.5, 73.3, 72.6, 56.0, 55.0, 20.4 ppm. MS (EI⁺): m/e 534 (M⁺), 459, 443, 415, 383, 311, 301, 211, 91. HRMS (EI+): calcd for C₃₁H₃₄O₈ (M⁺), *m/e* 534.2254; found, *m/e* 534.2243.

1-O-Acetyl-3-O-(p-methoxybenzyl)-4,5-O-dibenzyl-2deoxyl-2-oxo-myo-inositol (9a). To a solution of enol acetate 8a (2.20 g, 4.12 mmol) in 100 mL of acetone-water (3:2) was added mercuric acetate (13.20 g, 41.2 mmol); the solution was stirred for 45 min, 57 mL of saturated NaCl solution was added, and the mixture was stirred for 24 h. The acetone was evaporated, and the residue was extracted (EtOAc), concentrated, and purified on SiO₂ (30-50% EtOAc-hexane) to give 1.85 g of inositol 9a (86% yield, 90% pure): ¹H NMR (250 MHz, CDCl₃) δ 7.40–7.10 (m, 12H, Bn + PMB), 6.86, 6.83 (2s, 2H, PMB), 5.12 (bs, 1H), 4.86 (d, J = 11.1 Hz, 2H), 4.75 (d, J =10.8 Hz, 2H), 4.64 (q, J = 11.4 Hz, 2H), 4.47 (d, J = 11.1 Hz, 1H), 4.25 (s, 1H), 4.09 (m, 1H), 3.80-3.70 (m, 1H), 3.74 (s, 3H, OMe), 2.17 (s, 3H, Ac) ppm; 13 C NMR (75 MHz, CDCl₃) δ 169.9, 159.9, 137.3, 129.8-128.1 (m), 114.1, 113.9, 83.5, 81.8, 78.5, 76.1, 74.9, 73.5, 73.1, 69.4, 55.3, 45.7 ppm. MS (CI⁺, NH₃): m/e 538 (MNH4⁺), 494, 461, 429, 418, 211, 181, 154, 137, 121, 108, 91. HRMS (CI⁺, NH₃): calcd for $C_{30}H_{36}NO_8$ (MNH₄⁺), m/e538.2441; found, m/e 538.2457.

1-O-Acetyl-4-O-(p-methoxybenzyl)-3,5-O-dibenzyl-2deoxyl-2-oxo-myo-inositol (9b). To a solution of enol acetate **8b** (3.0 g, 5.6 mmol) in 200 mL of acetone-water (3:2) was added mercuric acetate (18.6 g, 58.3 mmol). The solution was stirred for 45 min, and 100 mL of saturated NaCl solution was added. The mixture was stirred for 24 h and concentrated in vacuo, and the residue was extracted (3 volumes of CHCl₃) and purified by flash chromatography (33% EtOAc-hexane) to give 1.7 g of protected inositol **9b** (57% yield), $R_f \sim 0.36$ (50% EtOAc-hexane): ¹H NMR (200 MHz, CDCl₃) δ 7.41-7.23 (m, 12H, Bn + PMB), 6.89, 6.84 (d, J = 8.6 Hz, 2H, PMB), 5.17 (d, J = 2.4 Hz, 1H), 4.98-4.52 (m, 6H), 4.34 (m, 1H), 4.20-4.04 (m, 2H), 3.87 (m, 1H), 3.82 (s, 3H, OMe), 2.73 (s, 1H, OH), 2.23 (s, 3H, Ac) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 197.7, 169.7, 159.1, 137.2, 130.2-127.7 (m), 113.6, 106.5, 105.5, 83.4, 81.3, 78.7, 75.7, 74.7, 73.4, 73.2, 69.1, 55.1, 20.3 ppm. MS (FAB): $\mathit{m/e}\,519~(M-H^+)\,429,\,399,\,386,\,273,\,211,\,121.$ HRMS (FAB): calcd for $C_{30}H_{34}O_8~(M-H^+),~\mathit{m/e}\,519.2019;$ found, $\mathit{m/e}\,519.2020.$

1-O-Acetyl-3-O-(p-methoxybenzyl)-4,5-O-dibenzyl-myoinositol (10a). To a solution of inosose 9a (1.85 g, 3.56 mmol) in 60 mL of dry acetonitrile was added NaBH(OAc)₃ (7.55 g, 35.6 mmol) and 19 mL of glacial HOAc. The mixture was stirred for 45 min at room temperature, and the excess NaBH-(OAc)₃ was destroyed by dropwise addition of 0.5 M NaHSO₄. The mixture was extracted with EtOAc, washed (0.5 M NaHSO₄, saturated Na₂HPO₄), dried (Na₂SO₄), and concentrated. The residue was recrystallized from EtOAc/H to yield 1.5 g of **10a** (81%), $R_f \sim 0.21$ (67% EtOAc-hexane): ¹H NMR (300 MHz, CDCl₃) δ 7.40-7.10 (m, 12H, aromatic), 6.78 (m, 2H, PMB), 5.20-4.37 (m, 7H), 4.20 (m, 1H), 4.10-3.41 (m, 6H), 3.74 (s, 3H, OMe), 2.10 (s, 3H, Ac) ppm; $^{13}\mathrm{C}$ NMR (63 MHz, CDCl₃) & 172.3, 158.7, 130.0, 129.6, 128.1, 128.0, 113.8, 113.6, 82.6, 80.6, 80.1, 75.5, 72.9, 67.1, 55.8 ppm. MS (CI⁺, NH₃): m/e 540 (MNH₄⁺), 431, 401, 330, 311, 295, 277, 205, 181, 137, 121, 108, 91. HRMS: calcd for $C_{30}H_{38}NO_8$ (MNH₄⁺), m/e540.2597; found, m/e 540.2596.

1-O-Acetyl-4-O-(p-methoxybenzyl)-3,5-O-dibenzyl-myoinositol (10b). To a solution of inosose 9b (970 mg, 1.8 mmol) in 50 mL of dry acetonitrile was added NaBH(OAc)₃ (4.0 g, 18.8 mmol) and 8.8 mL of glacial HOAc. The mixture was stirred for 45 min at room temperature, and the excess NaBH-(OAc)₃ was destroyed by dropwise addition of 0.5 M NaHSO₄. The mixture was extracted (EtOAc), washed (0.5 M NaHSO₄), dried (Na_2SO_4), and purified on deactivated SiO_2 to give 800 mg of compound 10b (83% yield): ¹H NMR (200 MHz, CDCl₃) δ 7.34–7.22 (m, 12H, Bn + PMB), 6.85 (d, J = 8.6 Hz, 2H, PMB), 4.99-4.70 (m, 7H), 4.28 (t, J = 2.6 Hz, 1H), 4.10 (t, J = 9.4 Hz, 1H), 3.80 (s, 3H, OMe), 3.55 (dd, J = 2.6, 9.4 Hz, 1H), 3.36 (t, J = 9.4 Hz, 1H), 2.42 (s, 1H, OH), 2.22 (s, 1H, OH), 2.16 (s, 3H, Ac) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 170.8, 129.4-127.7 (m), 113.5, 108.5, 106.5, 83.0, 80.5, 80.0, 76.3, 75.5, 73.1, 73.0, 70.1, 68.0, 55.1, 21.1 ppm. HRMS (FAB): calcd for C₃₀H₃₄O₈ (M⁺), m/e 522.2254; found, m/e 522.2267.

3-O-(p-Methoxybenzyl)-4,5-O-dibenzyl-2,6-O-bis(benzoxymethyl)-myo-inositol (12a). A mixture of protected inositol 10a (1.50 g, 2.87 mmol), proton sponge (1.5 g), n-Bu₄-NBr (100 mg), and benzoxyl chloromethyl ether (BOMCl, 2 mL) in 40 mL of dry acetonitrile was stirred overnight at room temperature, then at 35 °C for 10 h, and then at 55 °C overnight. TLC (product $R_f \sim 0.57$, 33% EtOAc-hexane) showed no starting material. The mixture was concentrated 3-fold, diluted with water, and extracted with EtOAc, and the extracts were washed (water), dried (MgSO₄), and concentrated to give oil 11a. The crude material 11a was dissolved in 50 mL of MeOH, 0.35 M NaOH (20 mL) was added, and the mixture was refluxed 2 h. The usual workup gave 1.52 g of protected inositol 12a (76%): ¹H NMR (300 MHz, CDCl₃) δ 7.40-7.10 (m, 22H), 6.90-6.70 (m, 2H, PMB), 5.00-4.50 (m, 15H), 4.20-4.10 (m, 1H), 3.96 (t, J = 9 Hz, 1H), 3.88-3.80(m, 2H), 3.76 (s, 3H, OMe), 3.43 (m, 2H) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 138.7, 130.1–127.0 (m), 113.8, 113.7, 96.6, 95.9, 83.0, 82.8, 81.7, 79.6, 75.8, 72.3, 71.1, 70.1, 69.8, 65.3, 55.3 ppm. MS (FAB): m/e 743 (MNa⁺), 599, 491, 371, 227, 211, 181, 176, 121. HRMS: calcd for C44H48NaO9 (MNa⁺), m/e 743.3196; found, m/e 743.3192.

1-*O*-Acetyl-3,5-*O*-dibenzyl-4-*O*-(*p*-methoxybenzyl)-2,6-*O*-bis(benzoxymethyl)-*myo*-inositol (11b). A mixture of inositol 10b (800 mg, 1.5 mmol), proton sponge (830 mg), *n*-Bu₄-NBr (25 mg), and BOMCI (0.86 mL) in 40 mL of dry acetonitrile was stirred overnight at room temperature, at 35 °C for 10 h, and then 55 °C overnight. The mixture was concentrated 3-fold, diluted with water, and extracted with EtOAc, and the extracts were washed (water), dried (MgSO₄), concentrated, and purified on SiO₂ to give intermediate 11b as a colorless oil in 57% yield: ¹H NMR (CDCl₃, 200 MHz) δ 7.36–7.18 (m, 22H, Bn + PMB), 6.81 (d, *J* = 8.7 Hz, 2H, PMB), 5.04–4.70 (m, 14H), 4.56 (t, *J* = 9.6 Hz, 1H), 4.41–4.39 (m, 1H), 4.27 (t, *J* = 10.0 Hz, 1H), 4.06 (t, *J* = 9.5 Hz, 1H), 3.80 (s, 3H, OMe), 3.59–3.48 (m, 2H), 1.85 (s, 3H, Ac) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 170.3, 159.0, 138.3, 137.7, 137.5, 129.5–127.2 (m), 113.5, 95.7, 95.0, 83.1, 81.0, 80.0, 76.3, 75.6, 75.4, 72.8, 72.6, 69.4, 69.1, 55.0, 20.7 ppm. HRMS (FAB): calcd for $C_{46}H_{50}O_{10}$ (M - H⁺), *m/e* 761.3326; found, *m/e* 761.3355.

1-O-Acetyl-3,5-O-dibenzyl-2,6-O-bis(benzoxymethyl)myo-inositol (12b). To the intermediate 11b (660 mg, 0.866 mmol) in 30 mL of wet CH₂Cl₂ was added DDQ (231 mg, 1.01 mmol). The mixture was stirred at room temperature for 3 h, diluted to 60 mL CH₂Cl₂, washed ($2 \times 10\%$ aqueous NaHCO₃, brine, water), dried (MgSO₄), concentrated, and purified on SiO₂ (25% EtOAc-hexane) to give 4-hydroxy compound 12b (500 mg, 89%) as a colorless oil, $R_f \sim 0.3$ (25% EtOAchexane): ¹H NMR (200 MHz, CDCl₃) δ 7.37-7.26 (m, 20H, Bn), 5.03-4.52 (m, 13H), 4.41 (t, J = 2.4 Hz, 1H), 4.23 (t, J = 8.8 Hz, 1H), 4.14 (t, J = 9.6 Hz, 1H), 3.49-3.35 (m, 2H), 2.52 (s, 1H, OH), 1.85 (s, 3H, Ac); 13 C NMR (50 MHz, CDCl₃) δ 170.3, 138.3, 137.6, 137.2, 128.4-127.3 (m), 107.5, 105.5, 95.7, 94.7, 82.8, 79.1, 75.1, 72.7, 72.1, 71.7, 69.5, 69.1, 20.7 ppm. HRMS (FAB): calcd for C₃₈H₄₂O₉ (MH⁺): *m*/*e* 643.2907; found, m/e 643.2926.

4,5-*O*-**Dibenzyl-2,6**-*O*-**bis(benzoxymethyl)**-*myo*-inositol **(13a).** A mixture of protected inositol **12a** (40 mg, 0.056 mmol) and DDQ (50 mg) in 10 mL of $CH_2Cl_2-H_2O$ (100/1 v/v) was stirred at room temperature for 2 h. The usual workup gave 22 mg (67% yield) of diol **13a** ($R_f \sim 0.36$, 1.5:1 EtOAc-hexane); ¹H NMR (300 MHz, CDCl₃) δ 7.50–7.00 (m, 22H, aromatic), 5.00–4.50 (m, 12H), 4.25–4.00 (m, 1H), 3.85–3.30 (6H), 3.00 (br, s, 1H) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 138.6–137.4 (m), 129.0–128.0 (m), 82.4, 77.2, 75.5, 75.4, 71.8, 71.0, 70.3, 70.1, 64.8 ppm. MS (FAB): *m/e* 623 (MNa⁺).

1-O-Acetyl-3,5-O-dibenzyl-4-O-(dibenzylphosphoryl)-2,6-O-bis(benzoxymethyl)-myo-inositol (13b). To 4-alcohol 12b (100 mg, 0.159 mmol) and 1H-tetrazole (134 mg, 1.91 mmol) in 10 mL of dry CH₂Cl₂ was added a solution of dibenzyl N,N-diisopropylphosphoramidite (132 mg, 0.38 mmol). The solution was stirred at room temperature under nitrogen for 1 h and cooled to -40 °C, and a solution of *m*-CPBA (83 mg, 0.65 mmol) was added. The mixture was stirred for 1 h at room temperature, diluted with 50 mL of CH₂Cl₂, washed (10% aqueous NaHCO₃, brine), dried (MgSO₄), concentrated, and purified on SiO₂ (50% EtOAc-hexane) to give 100 mg (71%) of product **13b** as a colorless oil, $R_f \sim 0.72$ (50% EtOAchexane): ¹H NMR (200 MHz, CDCl₃) δ 7.34-7.04 (m, 30H, Bn), 4.98–4.59 (m, 17H), 4.49 (J = 12.0 Hz, 1H), 4.37 (m, 1H), 4.27 (t, J = 12.0 Hz, 1H), 3.60–3.50 (m, 2H), 1.81 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.5, 138.1, 137.7, 137.1, 128.4-127.4 (m), 106.7, 105.7, 104.7, 96.0, 95.0, 81.8, 81.7, 79.6, 79.5, 78.0, 75.0, 72.4, 72.3, 72.0, 69.7, 69.3, 69.2, 69.1, 69.0, 20.7 ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.56 ppm. HRMS (FAB): calcd for C₅₂H₅₅O₁₂P (M⁺), *m/e* 902.3431; found, *m/e* 902.3480.

Tetrabenzyl 1,3-(4,5-O-dibenzyl-2,6-O-bis(benzoxymethyl)-myo-inosityl) Bis(phosphate) (14a). A solution of diol 13a (20 mg, 0.038 mmol) in 2 mL of CH₂Cl₂ and 1H-tetrazole (10 mg, 0.14 mmol) was stirred at room temperature, while a solution of dibenzyl N,N-diisopropylphosphoramidite (40 mg, 0.12 mmol) in 1 mL of CH₂Cl₂ was added. The mixture was stirred at room temperature for 1 h and cooled to -40 °C, m-CPBA (20 mg) was added, and the reaction was stirred for 30 min at 0 °C and then 30 min at room temperature. The mixture was diluted with CH₂Cl₂, washed (10% aqueous NaHCO₃), dried (Na₂SO₄), concentrated, and chromatographed on SiO₂ using 50% EtOAc-hexane to give 40 mg (87%) of protected bis(phosphate) 14a as a viscous colorless oil, $R_f \sim 0.3$: ¹H NMR (300 MHz, CDCl₃) δ 7.4–7.2 (m, 40H, phenyl), 5.0-4.4 (m, 22H), 4.4-4.1 (m, 2H), 4.0 (m, 1H), 3.4 (m, 1H) ppm; 13 C NMR (63 MHz, CDCl₃) δ 138.1, 137.8, 135.7, 130-127 (m), 96.1, 75-74 (m), 76-69 (m) ppm; ³¹P NMR (101 MHz, CDCl₃) δ 0.33, 0.09 (1:1) ppm.

1,3-*myo*-Inosityl Bis(phosphate) Tetrasodium Salt (15). A solution of bis(phosphate) **14a** (140 mg, 0.125 mmol), NaHCO₃ (42 mg, 0.5 mmol), and Pd/C (10%, 50 mg) in *t*-BuOH-H₂O (6:1 v/v, 35 mL) was shaken at 50 psi initial pressure for 5 h. The catalyst was filtered off and washed with 2×5 mL of EtOH, 2×5 mL of EtOH-H₂O (1:1 v/v), and 2×5 mL of water. Removal of solvent gave 50 mg of solid bis-(phosphate) **15**: ¹H NMR (200 MHz, D₂O) δ 4.27 (t, J = 2.8

Hz, 1H), 4.0–3.80 (m, 2H), 3.72 (t, J = 9.2 Hz, 2H), 3.35 (t, J = 9.2 Hz, 1H) ppm; ³¹P NMR (81 MHz, D₂O) δ 6.70, 5.0 (1:1) ppm.

Cyanoethyl 1,2-O-Dipalmitoyl-sn-glyceryl 1-[3-(p-Methoxybenzyl-4,5-O-dibenzyl)-2,6-O-bis(benzoxymethyl)-myoinosityl] Phosphate (17a). To a mixture of protected inositol 12a (400 mg, 0.556 mmol) and 1H-tetrazole (155 mg, 2.21 mmol) in 5 mL of dry CH₂Cl₂ was added phosphoramidite 16 (840 mg, 1.09 mmol) in 2 mL of CH₂Cl₂, and the mixture was stirred at room temperature for 30 min. The mixture was cooled to -40 °C, and m-CPBA (250 mg, 60-85%) was added and stirred as it warmed to room temperature. The mixture was diluted to 20 mL with CH₂Cl₂, washed (10% NaHCO₃, water), dried (Na₂SO₄), concentrated, and chromatographed on SiO $_2$ using 50% EtOAc-hexane to give 760 mg (97%) of protected phosphoinositide **17a** ($R_f \sim 0.3$) as a viscous, colorless oil: ¹H NMR (250 MHz, CDCl₃) δ 7.35-7.20 (m, 22H, aromatic), 6.80-6.60 (m, 2H, PMB), 5.30-5.10 (m, 1H), 5.05-4.40 (m, 14H), 4.35-4.15 (m, 10H), 3.78 (s, 3H, OMe), 3.80-3.30 (m, 2H), 2.52 (J = 6.1 Hz, CH₂CN), 2.28 (t, J = 7.2 Hz, 4H, COCH₂), 1.70-1.50 (m, 4H), 1.30-1.10 (m, 48H), 0.90 (t, J = 4.8 Hz, 6H, 2 \times CH₃) ppm; ¹³C NMR (63 MHz, CDCl₃) δ 171.1, 133.4, 129.4-127.6 (m), 113.8, 95.5, 87.5, 82.8, 81.4, 75.7, 72.6, 70.3, 69.8, 69.7, 61.5, 55.2, 34.1, 34.0, 29.7-29.1 (m), 24.8, 22.7, 22.5, 14.3 ppm; ³¹P NMR (81 MHz, CDCl₃) δ -0.44, -0.63 (1:1) ppm.

3,5-O-Dibenzyl-4-O-dibenzylphosphoryl-2,6-O-bis(benzoxymethyl)-myo-inositol (14b). To a solution of acetate 13b (200 mg, 0.221 mmol) in 20 mL of methanol was added K₂CO₃ (61.3 mg, 0.443 mmol), and the mixture was stirred at room temperature for 45 min until TLC showed that the reaction was complete. The solvent was evaporated, and the residue was dissolved in water and extracted with $3 \times$ with EtOAc. The combined organics were washed (water, brine), and dried (MgSO₄), and the crude product was purified on SiO₂ with 4% EtOAc-CH₂Cl₂ to give 130 mg of **14b** (68%) ($R_f \sim 0.48$, 50% EtOAc-hexane): ¹H NMR (200 MHz, CDCl₃) δ 7.38-7.08 (m, 30H, aromatic), 5.02-4.52 (m, 17H), 4.26 (s, 1H), 3.94-3.84 (m, 2H), 3.56-3.45 (m, 2H) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 139.2–136.8 (m), 128.3–127.1 (m), 96.4, 95.5, 82.8, 81.1, 79.7, 79.5, 74.8, 74.7, 71.7, 70.8, 70.0, 69.5, 69.0, 68.9, 66.8 ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.56 ppm. HRMS (FAB): calcd for C₅₀H₅₄O₁₁P (MH⁺), *m*/*e* 861.3404; found, *m*/*e* 861.3438

Cyanoethyl 1,2-O-Dipalmitoyl-sn-glyceryl 3,5-O-dibenzyl-4-O-dibenzylphosphoryl-2,6-O-dibenzoxymethyl-myoinositol Phosphate (17b). To a mixture of inositol 14b (43 mg, 0.050 mmol) and 14 mg (0.20 mmol) of 1H-tetrazole in dry CH₂Cl₂ (10 mL) was slowly added cyanoethyl 1,2-Odipalmitoyl-sn-glyceryl N,N-diisopropylphosphoramidite (16) (150 mg, 0.20 mmol) in CH₂Cl₂ (1 mL), and the mixture was stirred for 1 h at room temperature. The reaction was cooled to -40 °C, and m-CPBA (43 mg, 0.25 mmol) was added and stirred as it warmed to room temperature for another 1 h. Workup as above gave phosphate 17b (28 mg, 36% yield) as a colorless oil that slowly solidified: ¹H NMR (200 MHz, CDCl₃) δ 7.39–7.00 (m, 30H, Bn), 5.31–5.13 (m, 1H), 5.00–4.48 (m, 17H), 4.26-4.09 (m, 9H), 3.67-3.48 (m, 2H), 2.51-2.45 (t, J = 6.2 Hz, 2H), 2.28 (t, J = 7.0 Hz, 2H, CH₂CO), 2.27 (t, J = 7.2 Hz, 2H, CH₂CO), 1.80-1.50 (m, 4H), 1.25 (m, 48H), 0.95-0.91 (t, J = 6.2 Hz, 6H, CH₃) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 173.2, 172.8, 138.0, 137.7, 137.6, 137.3, 136.1, 136.0, 135.8, 128.3-127.3 (m), 116.5, 96.5, 95.3, 81.3, 79.5, 79.3, 75.0, 73.1, 72.4, 70.3, 69.6, 69.1-68.0 (m), 67.9, 66.0, 65.9, 62.3, 62.3, 61.5, 34.1, 33.9, 31.9, 29.7-29.1 (m), 25.5, 24.8, 22.7, 19.5, 19.4, 14.1 ppm; ³¹P NMR (81 MHz, CDCl₃) δ -0.47 (1 P), -1.09 (1 P) ppm. MS (FAB): m/e 1567 (MNa⁺), 551, 313, 181. HRMS (FAB): calcd for C₈₈H₁₂₃NNaO₁₈P₂ (MNa⁺), *m/e* 1566.8113; found, m/e 1566.8078.

The low yields (36–40%) in the coupling reactions to give **17b** and **34a** below do not reflect a fundamental limitation in the reaction. Instead, low recoveries simply reflect difficulties in workup, primarily due to (a) loss of material at foamy aqueous/organic interfaces and (b) irreversible adsorption to silica gel during purification. **Cyanoethyl 1,2-O-Dipalmitoyl-***sn***-glyceryl 1-(4,5-O-dibenzyl-2,6-O-bis(benzoxymethyl)***-myo***-inosityl) Phosphate (18).** A mixture of PMB ether **17a** (800 mg, ~90% pure, 0.556 mmol) and DDQ (150 mg, 0.661 mmol) was stirred in 50 mL of CH₂Cl₂-H₂O (100:1, v:v) at room temperature for 2 h. The mixture was washed (10% aqueous NaHCO₃), dried (MgSO₄), and purified on SiO₂ using 67% EtOAc-hexane to give 430 mg of phosphate **18** (60% yield from **12a**) as an oil, $R_i \sim 0.34$: ¹H NMR (250 MHz, CDCl₃) δ 7.35-7.20 (m, 20H, aromatic), 5.30-5.10 (m, 1H), 5.05-4.40 (m, 12H), 4.35-4.15 (m, 10H), 3.80-3.30 (m, 2H), 3.00 (br, 1H, OH), 2.52 (*J* = 6.1 Hz, CH₂CN), 2.28 (t, *J* = 7.2 Hz, 4H, COCH₂), 1.70-1.50 (m, 4H), 1.30-1.10 (m, 48H), 0.90 (t, *J* = 4.8 Hz, 6H, 2 × CH₃) ppm; ³¹P NMR (81 MHz, CDCl₃) δ -0.63, -0.81 (1:1) ppm.

Cyanoethyl 1,2-O-Dipalmitoyl-sn-glyceryl 1-(3-O-Dibenzylphosphoryl-4,5-O-dibenzyl-2,6-O-bis(benzoxymethyl)myo-inosityl) Phosphate (19). A solution of hydroxy compound 18 (430 mg, 0.335 mmol) in 5 mL of CH₂Cl₂ and 1Htetrazole (95 mg, 1.36 mmol) was stirred at room temperature, while a solution of dibenzyl N,N-diisopropylphosphoramidite (230 mg, 0.67 mmol) in 1 mL of CH_2Cl_2 was added. The mixture was stirred at room temperature for 1 h and cooled to -40 °C, m-CPBA (115 mg) was added, and the reaction was stirred for 30 min at 0 °C and then 30 min at room temperature. The mixture was diluted with CH_2Cl_2 and washed (10%) NaHCO₃, water), dried (Na₂SO₄), concentrated, and chromatographed on SiO₂ using 50% EtOAc-hexane to give 414 mg (80%) of compound **19** ($R_f \sim 0.38$) as a viscous colorless oil: ¹H NMR (300 MHz, CDCl₃) δ 7.4–7.2 (m, 40H, phenyl), 5.0– 4.4 (m, 22H), 4.4-4.1 (m, 2H), 4.0 (m, 1H), 3.4 (m, 1H) ppm; ³¹P NMR (81 MHz, CDCl₃) & 0.33, 0.09 (1:1) ppm. MS (FAB): m/e 1567 (MNa⁺), 808, 553, 552, 550, 313, 281, 267, 221, 207, 147, 136, 133. HRMS (FAB): calcd for C888H123NNaO18P2 (MNa⁺), m/e 1566.8113; found, m/e 1566.8160.

3-Aminopropyl 1,2-*O*-Dipalmitoyl-*sn*-glyceryl 1-(3-*O*-Phosphoryl-*myo*-inosityl) Phosphate (20a). A solution of fully protected precursor 19 (170 mg, 0.11 mmol), NaHCO₃ (9 mg, 0.10 mmol), and Pd/C (10%, 100 mg) in *t*-BuOH-H₂O (6: 1, v:v, 35 mL) was added, and the mixture was shaken at 50 psi initial pressure for 5 h. The catalyst was filtered off and washed with 3×5 mL of EtOH, 5 mL of EtOH-H₂O (v/v 1:1), and 1×5 mL of water. Removal of solvent gave 52 mg (63% yield) of solid PtdIns(3)P triester 20a: ¹H NMR (200 MHz, D₂O) δ 4.30–3.20 (m, 13H), 2.70 (m, 2H), 2.30–2.20 (m, 4H, COCH₂), 1.7–1.4 (m, 6H), 1.30–1.00 (m, 48H), 0.90–0.70 (m, 6H) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 8.8 (s and sharp), 1.9 (br) (~ 1:1) ppm.

3-Aminopropyl 1,2-*O*-Dipalmitoyl-*sn*-glyceryl 4-*O*-Phosphoryl-D-*myo*-inositol Phosphate Sodium Salt (20b). A mixture of cyanoethyl ester 17b (43 mg, 0.028 mmol), sodium bicarbonate (2.4 mg, 0.028 mmol), and Pd/C (10%) (100 mg) in 14 mL of *t*-BuOH/water (6:1) was shaken under hydrogen (50 psi) for 10 h and filtered, the catalyst was washed with a CHCl₃-CH₃OH-H₂O mixture, and the filtrate was concentrated under N₂ to give PtdIns(4)P triester **20b**: ¹H NMR (200 MHz, D₂O) δ 5.24 (s, 1H), 5.00-2.60 (br), 2.40-2.14 (m, 4H), 1.77-1.50 (m, 4H), 1.47-1.04 (m, 48H), 0.95-0.84 (m, 6H); ³¹P NMR (81 MHz, CDCl₃) δ 0.09 (s), 2.6 (s) ppm.

1,2-O-Dipalmitoyl-*sn***-glyceryl 1-[3-O-Phosphoryl-***myo***-inosityl] Phosphate (21).** A solution of precursor **19** (130 mg, 0.086 mmol) and *i*-Pr₂NEt (200 μ L) in 2 mL of MeOH was stirred overnight (TLC showed no remaining starting material) and evaporated to dryness. The residue was dissolved in *t*-BuOH–H₂O (6:1 v/v, 35 mL) and NaHCO₃ (10 mg, 0.11 mmol), Pd/C (10%, 100 mg) was added, and the mixture was shaken at 50 psi initial pressure for 5 h. The catalyst was filtered off and washed with 5×5 mL of EtOH, 3×5 mL of EtOH–H₂O (v/v 1:1), and 2×5 mL of water. Removal of solvent gave 62 mg (77% yield) of solid PtdIns(3)P di-C₁₆diester **21**: ¹H NMR (200 MHz, CDCl₃) δ 5.30–3.50 (very broad and poorly resolved peaks, 11H), 2.30–2.10 (m, 4H), 1.70–1.30 (m, 52H), 0.90–0.80 (m, 6H) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 3.8, 0.1 (~1:1) ppm.

In the processing of this and other hydrogenolytic deprotection reactions of these detergent-like molecules, an estimated equimolar (according to the number of negative phosphate charges in the molecule) amount of NaHCO₃ was generally added during hydrogenation to suppress acyl migration. When traces of Na⁺ salts remain, the ³¹P NMR peaks become complex as the different counterions result in different chemical shifts. Thus, the hydrogenolysis reaction mixture was first passed through a Dowex 50WX8-200 ion-exchange resin and concentrated to dryness to remove excess HCO_3^- . It was then passed through a Chelex column to exchange the cation back to Na⁺. When NaHCO₃ was not used, this extra step was unnecessary.

3-((p-Benzoyldihydrocinnamyl)amino)propyl 1,2-O-Dipalmitoyl-sn-glyceryl 1-(3-O-Phosphoryl-myo-inosityl) Phosphate (22a). PtdIns(3)P aminopropyl triester 20a (5 mg, 0.0056 mmol) was suspended in 2.5 mL of anhydrous DMF, and N-hydroxysuccinimido p-benzoyldihydrocinnamate ester (BZDC-NHS ester, 3.6 mg, 0.010 mmol) was added, followed by 0.2 mL of dry Et₃N. The suspension was stirred overnight and concentrated in vacuo, and the residue was evaporated with 2 mL of water. The residue was centrifuged with acetone several times until the acetone showed no remaining UVabsorbing material. Then the solid was dissolved in 1 mL of water and applied to a 50×6 mm column of DEAE cellulose (OH⁻ form). The column was washed with 2×1 mL of water and then eluted with 2 \times 1 mL of 0.1 M TEAB, 2 \times 1 mL of 0.2 M TEAB, 2 \times 1 mL of 0.3 M TEAB, and 1 mL of 0.4 M TEAB buffer. The product eluted in 0.2 M buffer. Evaporation of these fractions in vacuo followed by evaporating with several small volumes of methanol gave the triethylamine salt of BZDC-PtdIns(3)P triester 22a. After ion-exchange using a column packed with Bio-Rad Chelex 100 resin (Na form), the sodium salt of 22a (2.5 mg, 40% yield) was obtained as a glass: ¹H NMR (250 MHz, D₂O) δ 7.86–7.75 (m, 5H), 7.50 (t, 2H), 7.36 (t, 2H), 4.30-3.20 (m, 13H), 3.10 (m, 2H), 2.90 (t, J = 7.5 Hz, 2H), 2.45 (t, J = 7.5 Hz, 2H), 2.30–2.20 (m, 4H, COCH₂), 1.7-1.4 (m, 6H), 1.30-1.00 (m, 48H), 0.90-0.70 (m, 6H) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 8.8 (s), 1.9 (s) (~1:1) ppm.

3-((p-Benzoylditritiocinnamyl)amino)propyl 1,2-O-Dipalmitoyl-sn-glyceryl 1-[3-O-Phosphoryl-myo-inosityl] **Phosphate (22a').** $[{}^{3}H_{2}]$ -*N*-Hydroxysuccinimido *p*-benzoyldihydrocinnamate ester ([3H]BZDC-NHS ester, 2 mCi, 40 Ci/ mmol) in EtOAc was gently concentrated with dry N₂, and then 3-aminopropyl triester 20a (800 µg/mL of DMF stock solution, 15 μ L) was added followed by 10 mL of dry triethylamine. The suspension was stirred overnight. The solvents were evaporated in vacuo, the residue was evaporated with 1 mL of water, and the residue was dissolved in 1 mL of water and then applied to 50 \times 6 mm column of DEAE cellulose (OH⁻ form). The column was washed with 2×1 mL of water and then eluted with 2×1 mL of 0.1 M TEAB, 2×1 mL of 0.2 M TEAB, $2\,\times\,1$ mL of 0.3 M TEAB, and 1 mL of 0.4 M TEAB buffer. The product 22a' eluted in 0.1 M buffer. Radioactive fractions were pooled to give ca. 150 µCi of purified 22a', specific activity 42.5 Ci/mmol (radiochemical yield, 7.5%).

3-((*p*-Benzoyldihydrocinnamyl)amino)propyl 1,2-*O*-Dipalmitoyl-*sn*-glyceryl 1-(4-*O*-phosphoryl-*myo*-inosityl) Phosphate (22b). To aminopropyl triester 20b (5 mg, 0.0056 mmol) in 2 mL of DMF (distilled after incubation with P₂O₅ for 5 days) was added BZDC-NHS ester (3.6 mg, 0.010 mmol), and the mixture was stirred for 30 min. Dry Et₃N (0.2 mL) was then added, and the suspension was stirred overnight. DMF was removed under a stream of N₂, and the residue was washed with acetone (5×1 mL) to give 22b as a white powder (2.3 mg, 37% yield), which was homogeneous by TLC: ¹H NMR (250 MHz, D₂O) δ 7.83–7.71 (m, 5H), 7.64–7.54 (m, 2 H), 7.45–7.38 (m, 2H), 4.29–3.40 (m, 13H), 3.23–3.09 (m, 2H), 2.75–2.70 (m, 4H), 2.03–1.96 (m, 4H), 1.67–1.49 (m, 4H), 1.31–1.20 (m, 48H), 0.96–0.79 (m, 6H) ppm; ³¹P NMR (81 Hz, D₂O) 4.89 (br) ppm.

Benzyl 1,2- \hat{O} Diisopropylidene-*sn*-glyceryl *N*,*N*-Diisopropylphosphoramidite (23). A mixture of BnOP(NPr₂-i)₂ (800 mg, 2.36 mmol), 1,2-O-diisopropylidene-*sn*-glycerol (312 mg, 2.36 mmol), and diisopropylammonium tetrazole (200 mg) in 10 mL of dry CH₂Cl₂ was stirred at room temperature for 1

h and then washed (10% NaHCO₃), dried (MgSO₄), and purified on SiO₂ using EtOAc-hexane-Et₃N (100:300:10, v/v/ v), to give 732 mg of phosphoramidite **23** (84%), $R_f \sim 0.95$: ¹H NMR (200 MHz, CDCl₃) δ 7.40–7.20 (m, 5H, aromatic), 4.89 (d, J = 8.4 Hz, CH₂O), 4.80–4.50 (m, 1H), 4.30–4.10 (m, 2H), 4.01, 3.99 (2t, J = 8.2 Hz, 2H), 3.80–3.50 (m, 2H), 1.41, 1.35 (2s, 6H, isopropylidene) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 148.8, 148.7 (~1:1) ppm.

Benzyl 1,2-O-Diisopropylidene-sn-glyceryl 1-[3-(p-Methoxybenzyl)-4,5-O-dibenzyl-2,6-O-dibenzoxymethylmyo-inosityl] Phosphate (24). To a mixture of precursor 12a (200 mg, 0.28 mmol) and 1H-tetrazole (80 mg, 1.1 mmol) in 5 mL of dry CH₂Cl₂ was added phosphoramidite 23 (200 mg, 1.09 mmol) in 2 mL of CH₂Cl₂, and the mixture was stirred at room temperature for 30 min. The mixture was cooled to -40 °C, and *m*-CPBA (250 mg, 60–85%) was added and stirred as the reaction warmed to room temperature. The mixture was diluted to 20 mL with CH₂Cl₂, washed (10% NaHCO₃, water), dried (Na₂SO₄), concentrated, and chromatographed on SiO_2 using 50% EtOAc-hexane to give 240 mg (97%) of compound **24** as a viscous colorless oil ($R_f \sim 0.4$): ¹H NMR (200 MHz, CDCl₃) & 7.40-7.20 (m, 27H, aromatic), 6.80 (m, 2H, PMB), 5.20-4.50 (m, 14H), 4.30-3.90 (m, 8H), 3.78 (s, 3H, OMe), 3.70-3.40 (m, 6H), 1.37, 1.31 (2s, 6H, isopropylidene) ppm; ³¹P NMR (81 MHz, CDCl₃) δ:-0.19, -0.34 (~1:1) ppm. MS (FAB): m/e 1027 (MNa⁺), 669, 386, 370, 328, 211, 181, 121. HRMS (FAB): calcd for $C_{57}H_{65}NaO_{14}P$ (MNa⁺), m/e1027.4010; found, m/e 1027.4031.

Benzyl 1,2-*O*-Diisopropylidene-*sn*-glyceryl 1-[4,5-*O*-Dibenzyl-2,6-*O*-bis(benzoxymethyl)-*myo*-inosityl] Phosphate (25). A mixture of phosphate 24 (230 mg, 0.23 mmol) and DDQ (80 mg, 0.35 mmol) in 20 mL of $CH_2Cl_2-H_2O$ (100/1 v/v) was stirred at room temperature for 2 h. The mixture was washed (10% NaHCO₃), dried (MgSO₄), and purified on SiO₂ using 50% EtOAc-hexane to give 140 mg (69% yield) of phosphate 25 as an oil ($R_f \sim 0.20$): ¹H NMR (200 MHz, CDCl₃) δ 7.40–7.15 (m, 25H, aromatic), 5.20–4.50 (m, 12H), 4.30–3.90 (m, 7H), 3.80–3.40 (m, 6H), 3.30 (br., 1H, OH), 1.39, 1.32 (2s, 6H, isopropylidene) ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.45, –0.56 (~1:1) ppm. MS (FAB): *m/e* 907 (MNa⁺), 855 (MH⁺), 777, 669, 657, 579, 393, 271, 182, 181, 165, 154, 136, 115. HRMS: calcd for C₄₉H₅₇NaO₁₃P (MNa⁺), *m/e* 907.3435; found, *m/e* 907.3476.

Benzyl sn-glyceryl 1-(3-O-Dibenzylphosphoryl-4,5-Odibenzyl-2,6-O-dibenzoxymethyl-myo-inosityl) Phosphate (27). A solution of protected phosphoinositide 25 (136 mg, 0.154 mmol) in 2 mL of CH₂Cl₂ and 1H-tetrazole (43 mg, 0.61 mmol) was stirred at room temperature, while a solution of dibenzyl N,N-diisopropylphosphoramidite (110 mg, 0.32 mmol) in 1 mL of CH₂Cl₂ was added. The mixture was stirred at room temperature for 1 h and cooled to -40 °C, and *m*-CPBA (20 mg) was added. The reaction was stirred for 30 min at 0 °C and then 30 min at room temperature, and the mixture was diluted with CH₂Cl₂, washed (10% NaHCO₃, water), dried (Na₂SO₄), and concentrated to give a crude residue 26 (33% EtOAc-hexane, $R_f \sim 0.5$). This residue was dissolved in MeOH, 10 mg of *p*-TsOH monohydrate was added, and the mixture was stirred at room temperature for 1 h. The acid was neutralized (0.5 mL of 10% aqueous NaHCO₃), and the solvent was evaporated. Purification on SiO2 eluting with 67% EtOAc-hexane gave 100 mg ($R_f \sim 0.12$) of compound **27** (59% for two steps) as an oil: ¹H NMR (200 MHz, CDCl₃) δ 7.40-7.10 (m, 35H, aromatic), 5.20-4.50 (m, 18H), 4.45-3.50 (m, 11H), 2.0 (br, 2H, OH) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 0.98, $-0.64, 0.75, -1.09 (\sim \hat{1}:\hat{1}:1:1)$ ppm.

Benzyl 1,2-*O*-Diacyl-*sn*-glyceryl 1-(3-*O*-Dibenzylphosphoryl-4,5-*O*-dibenzyl-2,6-*O*-bis(benzoxymethyl)-*myo*-inosityl) Phosphate (28). A solution of diol 27 (1 equiv) and fatty acid (octanoic acid or butyric acid, 3 equiv), DCC (3 equiv), and DMAP (0.05-0.1 equiv) in 5 mL of dry CH₂Cl₂ was stirred overnight. The solid was removed by filtration, washed with ether, and concentrated, the residue was dissolved in 50% EtOAc-hexane, and the precipitate was removed by filtration and washed with the same solvent. Concentration of the filtrates gave an oil that was purified on SiO₂ using 50% EtOAc-hexane to give the phosphoinositides **28a** or **28b** with $R_f \sim 0.2$.

Data for benzyl 1,2-*O*-dioctanoyl-*sn*-glyceryl 1-(3-*O*-dibenzylphosphoryl-4,5-*O*-dibenzyl-2,6-*O*-bis(benzoxym-ethyl)-*myo*-inosityl) phosphate (28a): 40 mg yield; ¹H NMR (200 MHz, CDCl₃) δ 7.35–7.10 (m, 35H, aromatic), 5.15–4.00 (m, 27H), 3.60–3.40 (m, 2H), 2.2 (m, 4H, COCH₂), 1.70–1.20 (m, 20H), 0.9–0.7 (m, 6H, 2 × CH₃) ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.48, –0.53, –0.58, –0.64 (~1:1:1:1) ppm. MS (FAB): *m/e* 1380 (MNa⁺), 327, 255, 181, 127. HRMS (FAB): calcd for C₇₆H₉₄NaO₁₈P₂ (MNa⁺), *m/e* 1379.5813; found, *m/e* 1379.5759.

Data for benzyl 1,2-*O*-dibutyroyl-*sn*-glyceryl 1-(3-*O*-dibenzylphosphoryl-4,5-*O*-dibenzyl-2,6-*O*-bis(benzoxym-ethyl)-*myo*-inosityl) phosphate (28b): 50 mg yield; ¹H NMR (200 MHz, CDCl₃) δ 7.40–7.10 (m, 35H, aromatic), 5.20–4.50 (m, 20H), 4.40–3.90 (m, 7H), 3.60–3.40 (m, 2H), 2.30–2.20 (m, 4H, COCH₂), 1.70–1.50 (m, 4H), 1.00–0.80 (m, 6H, 2 × CH₃) ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.22, –0.45, –0.53 (~1:1:2) ppm. MS (FAB): *m/e* 1267 (MNa⁺), 215, 181. HRMS: calcd for C₆₈H₇₈NaO₁₈P₂ (MNa⁺), *m/e* 1267.4561; found, *m/e* 1267.4610.

1,2-O-Dioctanoyl-*sn***-glyceryl 1-(3-O-Phosphoryl-***myo***inosityl) Phosphate (29a).** To a solution of fully protected phosphate **28a** (50 mg, 0.037 mmol), *t*-BuOH–H₂O (6:1 v/v, 35 mL) and NaHCO₃ (9 mg, 0.10 mmol) was added Pd/C (10%, 50 mg), and the mixture was shaken at 50 psi initial pressure for 5 h. The catalyst was filtered off and washed with 5 mL of EtOH, 5 mL of EtOH–H₂O (v/v 1:1), and 3×5 mL of water. Removal of solvent gave 17 mg (63% yield) of solid PtdIns(3)P di-C₈-ester **29a**: ¹H NMR (200 MHz, D₂O) δ 5.35–5.15 (m, 1H), 4.45–4.20 (m, 2H), 4.10–3.60 (m, 7H), 3.35 (t, *J* = 9.2 Hz, 1H), 2.38, 2.34 (2t, *J* = 7.2 Hz, 4H, COCH₂), 1.7–1.2 (m, 20H), 0.82 (t, *J* = 6.2 Hz, 6H) ppm; ³¹P NMR (81 MHz, D₂O) δ 7.04, 2.65 ppm.

1,2-O-Dibutyryl-*sn***-glyceryl-1-(3-O-phosphoryl-***myo***inosityl) Phosphate (29b).** A solution of protected phosphoinositide **28b** (50 mg, 0.040 mmol), NaHCO₃ (10 mg, 0.11 mmol), and Pd/C (10%, 50 mg) in *t*-BuOH-H₂O (6:1 v/v, 35 mL) was shaken at 50 psi initial pressure for 5 h. The catalyst was filtered off and washed with 5 mL of EtOH, 5 mL of EtOH-H₂O (1:1 v/v), and 3×5 mL of water. Removal of solvent gave 22 mg (92%) of solid PtdIns(3)P di-C₄-ester **29b**: ¹H NMR (200 MHz, D₂O) δ 5.35-5.25 (m, 1H), 4.50-4.20 (m, 2H), 4.10-3.37 (m, 7H), 3.35 (t, J = 9.2 Hz, 1H), 2.37, 2.34 (2t, J = 7.2 Hz, 4H), 1.70-1.50 (m, 4H), 0.88, 0.86 (2t, J = 6.2 Hz, 6H, $2 \times$ CH₃) ppm; ³¹P NMR (81 MHz, D₂O) δ 7.43, 2.65 (~1:1) ppm.

General Procedure for the DCC/DMAP-Promoted Esterification. A mixture of a PMB-protected glycerol **30** (1 equiv), DMAP (0.05–0.1 equiv), fatty acid (2.5 equiv), and DCC (2.5 equiv) in dry CH_2Cl_2 (10 mL) was stirred at room temperature overnight. The mixture was filtered and concentrated. The residue was then dissolved in EtOAc, and the precipitate was filtered again and washed with EtOAc. The filtrate was concentrated and purified on SiO₂ column eluting with EtOAc-hexane mixtures to give the corresponding 1,2-O-diacyl-3-O-(p-methoxybenzyl)-sn-glycerol **31**.

Data for 1,2-*O***-dioctanoyl-3-***O***-(***p***-methoxybenzyl**)-*sn*glycerol (31b): obtained in 95% yield; ¹H NMR (200 MHz, CDCl₃) δ 7.24, 7.20, 6.88, 6.83 (4s, 4H, PMB), 5.28–5.17 (m, 1H), 4.46–4.45 (m, 2H), 4.31 (dd, J = 4.0, 12.0 Hz, 1H), 4.15 (dd, J = 6.4 Hz, 11.8 Hz, 1H), 3.79 (s, 3H, OMe), 3.54 (d, J =5.2 Hz, 2H), 2.30, 2.26 (2t, J = 7.8 Hz, 4H), 1.60–1.57 (m, 4H), 1.48–1.05 (m, 16H), 0.86 (t, J = 6.6 Hz, 6H, CH₃) ppm. MS (EI⁺): m/e 464 (M⁺) 352, 327, 226, 201, 135, 121. HRMS (EI⁺): calcd for C₂₇H₄₄O₆ (M⁺), m/e 464.3139; found, m/e464.3120.

Data for 1,2-*O***-dibutyryl-3-***O***-(***p***-methoxybenzyl**)*-sn***-glycerol (31c):** obtained in 90% yield; ¹H NMR (300 MHz, CDCl₃) δ 7.23, 7.19, 6.86, 6.84 (4s, 4H, PMB), 5.25–5.17 (m, 1H), 4.44 (AB, J = 11.7 Hz, 2H, OCH₂), 4.20 (d, AB, J = 3.8 Hz, 2H), 3.77 (s, 3H, OMe), 3.53 (d, J = 5.2 Hz, 2H), 2.26, 2.24 (2t, J = 7.4 Hz, 4H, COCH₂), 1.70–1.53 (m, 4H), 0.92,

0.91 (2t, J = 7.3 Hz, 6H, CH₃) ppm; ¹³C NMR (63 MHz, CDCl₃) δ : 173.2, 172.9, 159.3, 129.8, 129.3, 113.8, 87.7, 72.9, 70.0, 67.9, 62.7, 55.2, 36.2, 35.9, 18.4, 18.3, 13.6 ppm. MS (CI⁺): m/e 352 (M⁺) 281, 215, 145, 136, 121, 71. HRMS (CI⁺): calcd for C₁₉H₂₈O₆, m/e 352.1886; found, m/e 352.1881. Anal. Calcd for C₁₉H₂₈O₆: C, 64.77; H, 7.95. Found: C, 65.00, H, 7.88.

General Procedure for Deprotection of PMB Ethers. A mixture of a PMB ether **31** (1 equiv) in 20 mL of wet CH₂-Cl₂ (CH₂Cl₂-H₂O, 100:1 v/v) and DDQ (1.2 equiv) was stirred at room temperature for 3 h. It was then washed with 10% aqueous NaHCO₃ and brine, dried over MgSO₄, and concentrated. Purification on SiO₂ gel using 95% ether-CH₂Cl₂ mixtures gave the corresponding 1,2-*O*-diacyl-*sn*-glycerol **32**.

Data for 1,2-O-dioctanoyl-*sn***-glycerol (32b):** obtained in 80% yield; ¹H NMR (200 MHz, CDCl₃) δ 5.07 (quint, J = 5.6 Hz, 1H), 4.30–4.08 (m, 2H), 3.70 (d, J = 5.0 Hz, 2H), 3.37–2.27 (m, 4H), 1.66–1.54 (m, 4H), 1.28–1.20 (m, 16H), 0.89–0.83 (m, 6H) ppm. MS (CI+): *m/e* 362 (MNH₄⁺), 345, 327, 269, 226, 127, 98. HRMS (CI⁺): calcd for C₁₉H₄₀NO₅ (MNH₄⁺), *m/e* 362.2906; found, *m/e* 362.2909.

Data for 1,2-*O***-dibutyroyl***-sn***-glycerol (32c)** ($R_{f} \sim 0.35$, 33% EtOAc-hexane): obtained in72% yield; ¹H NMR (200 MHz, CDCl₃) δ 5.09 (quint., J = 5.0 Hz, 1H), 4.29, 4.26 (2q, J = 11.9 Hz, 2H), 3.72 (d, J = 4.9 Hz, 2H), 2.33, 2.30 (2t, J = 7.4 Hz, 4H, COCH₂), 1.70–1.53 (m, 4H), 0.95, 0.94 (2t, J = 7.2 Hz, 6H, CH₃) ppm. MS (CI⁺): 250 (MNH₄⁺), 233 (MH⁺), 215, 145, 88, 71. HRMS (CI⁺): calcd for C₁₁H₂₄NO₅ (MNH₄⁺), *m/e* 250.1654; found, *m/e* 250.1661.

General Procedure for Preparation of Phosphoramidites 33. A mixture of 1,2-*O*-diacyl-*sn*-glycerol and benzyl *N*,*N*-diisopropylchlorophosphoramidite and *N*,*N*-diisopropylethylamine was stirred at 0 °C for 2 h. It was washed with 10% aqueous NaHCO₃ solution and dried over MgSO₄. Purification on SiO₂ eluting with EtOAc-hexane-Et₃N (from 10: 50:3 to 10:30:3, v/v/v) gave the phosphoramidites 33. Spectroscopic data are shown; however, analytical data on these unstable materials were not obtained. These materials were employed directly for the coupling reactions to give adducts 34.

Data for benzyl 1,2-*O***-dioctanoyl**-*sn*-glycerol *N*,*N*-diisopropylphosphoramidite (33b): ¹H NMR (200 MHz, CDCl₃) δ 7.38–7.24 (m, 5H, Bn), 5.25–5.12 (m, 1H), 4.70 (t, *J* = 10.0 Hz, 2H), 4.40–4.31 (m, 1H), 4.23–4.10 (m, 1H), 3.81– 3.53 (m, 4H), 2.30 (t, *J* = 7.4 Hz, 4H), 1.67–1.56 (m, 4H), 1.27– 1.17 (m, 28H), 0.92–0.85 (m, 6H) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 149.6 (s), 149.4 (s) (1:1) ppm.

Data for benzyl 1,2-*O*-dibutyryl-*sn*-glycerol *N*,*N*-diisopropylphosphoramidite (33c): ¹H NMR (200 MHz, CDCl₃) δ 7.40–7.20 (m, 5H, Bn), 5.20–5.10 (m, 1H), 4.72 (d, *J* = 7.4 Hz, 2H), 4.40–4.20 (m, 2H), 3.80–3.50 (m, 4H), 2.30 (t, *J* = 7.2 Hz, 4H, COCH₂), 1.80–1.60 (m, 4H), 1.30 (m, 12H), 0.94 (2t, *J* = 7.4 Hz, 6H) ppm; ³¹P NMR (81 MHz, CDCl₃) δ 149.5 (s), 149.4 (s) (1:1) ppm.

Benzyl 1,2-O-Dipalmitoyl-sn-glyceryl 1-[3,5-O-Dibenzyl-2,6-O-dibenzoxymethyl-4-O-(dibenzylphosphoryl)-Dmyo-inositol] Phosphate (34a). To a mixture of alcohol 14b (40 mg, 0.047 mmol) and 1H-tetrazole (24 mg, 0.34 mmol) in CH₂Cl₂ (10 mL) was added a solution of benzyl 1,2-Odipalmitoyl-sn-glyceryl N,N-diisopropylphosphoramidite (33a, 190 mg) in 3 mL of dry CH₂Cl₂. Workup as in the preparation of 13b gave phosphate 34a (25 mg, 40% yield) as a colorless oil: ¹H NMR (200 MHz, CDCl₃) δ 7.38–6.99 (m, 35H, aromatic), 5.19-4.45 (m, 20H), 4.26-3.99 (m, 7H), 3.53-3.43 (m, 2H), 2.27-2.16 (m, 4H, CH2CO), 1.64-1.44 (m, 4H), 1.25 (m, 48H), 0.88 (t, 6H, J = 6.6 Hz, 6H, CH₃) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 137.9–135.8 (m), 128.4–127.0 (m), 107.5, 106.4, 105.5, 71.8, 69.9-66.7 (m), 61.3, 33.9-33.7 (m), 31.7, 29.5–28.9 (m), 24.6, 22.5, 13.9 ppm; ³¹P NMR (81 MHz, CDCl₃) $\delta = 0.22, -0.50 (\sim 1.1)$ ppm. MS (FAB): m/e 1604 (MNa⁺), 551, 313, 181. HRMS (FAB): calcd for C₉₂H₁₂₆NaO₁₈P₂ (MNa⁺), m/e 1603.8317; found, m/e 1603.8382.

Benzyl 1,2-*O*-dioctanoyl-*sn*-glyceryl 1-[3,5-*O*-dibenzyl-2,6-*O*-bis(benzoxymethyl)-4-*O*-(dibenzylphosphoryl)-D*myo*-inositol] phosphate (34b) was synthesized as described above: ¹H NMR (200 MHz, CDCl₃) & 7.39–7.01 (m, 35H, Bn), 5.15–4.45 (m, 20 H), 4.25–3.96 (m, 7H), 3.49–3.44 (m, 2H), 2.35–2.17 (m, 4H, CH₂CO), 1.70–1.45 (m, 4H), 1.26–1.24 (m, 16H), 0.88–0.84 (m, 6H, CH₃); ¹³C NMR (50 MHz, CDCl₃) δ 172.9, 172.5, 137.6–137.0 (m), 128.5–127.0 (m), 110.5, 109.5, 107.5, 106.5, 104.5, 102.5, 69.9–69.0 (m), 61.3, 33.8, 33.7, 31.4, 28.8–28.7 (m), 24.6, 22.4, 13.8 ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.20, –0.24 (combined as 1P), –0.53 (1P) ppm. MS (FAB): *m/e* 1380 (MNa⁺), 551, 327, 255, 181. HRMS (FAB): calcd for C₇₆H₉₄NaO₁₈P₂ (MNa⁺), *m/e* 1379.5813; found, *m/e* 1379.5795.

Benzyl 1,2-*O***-dibutanoyl**-*sn*-glyceryl 1-[3,5-*O*-dibenzyl-**2,6-***O*-bis(benzyloxymethyl)-4-*O*-(dibenzylphosphoryl)-D*myo*-inositol] phosphate (34c) was synthesized as described above: ¹H NMR (200 MHz, CDCl₃) δ 7.38–7.00 (m, 35H, Bn), 5.20–4.51 (m, 20H), 4.27–4.00 (m, 7H), 3.53–3.43 (m, 2H), 2.27–2.16 (m, 4H, CH₂CO), 1.66–1.53 (m, 4H), 0.95–0.85 (m, 6 H, CH₃) ppm; ¹³C NMR (50 MHz, CDCl₃) δ 172.7, 172.4, 137.9, 137.6, 137.5, 137.0, 128.5–127.1 (m), 112.5, 109.5, 108.5, 107.5, 106.5, 105.5, 104.5, 99.5, 69.9–66.9 (m), 61.3, 35.7, 35.6, 18.1, 13.4, 13.3 ppm; ³¹P NMR (81 MHz, CDCl₃) δ –0.19, –0.25 (combined as 1P), –0.53 (1P) ppm. HRMS (FAB): calcd for C₆₈H₇₈O₁₈P₂ (MH⁺), *m/e* 1245.4742; found, *m/e* 1245.4836.

1,2-O-Dipalmitoyl-*sn***-glyceryl 1-(4-O-Phosphoryl-D-***myo***-inositol) Phosphate Sodium Salt (35a).** A mixture of protected phosphoinositide **34a** (36 mg, 0.023 mmol), NaHCO₃ (6 mg, 0.072 mmol), and Pd/C (10%) (100 mg) in 12 mL of *t*-BuOH/water (6:1) was shaken under hydrogen (50 psi) for 14 h. The mixture was filtered, washed with a mixed solution of chloroform, methanol, and water, the solvents were evaporated, and the remaining product **35a** (17 mg, 84% yield) was lyophilized: ¹H NMR (200 MHz, CDCl₃) δ 5.20–3.70 (br), 3.20–2.50 (br), 2.20–1.90 (m, 4H), 1.55–1.35 (m, 4H), 1.35–

1.00 (m, 48H), 0.90–0.84 (m, 6H); $^{31}\mathrm{P}$ NMR (81 MHz, D2O) δ 7.67 (sharp), 2.92 (br) ppm.

1,2-O-Dioctanoyl-*sn***-glyceryl 1-(4-O-Phosphoryl-D-***myo***-inositol) Phosphate Sodium Salt (35b).** Preparation as above from protected phosphoinositide **34b** gave final product **35b** (80% yield) as the sodium salt: ¹H NMR (200 MHz, D₂O) δ 5.31–5.22 (m, 1H), 4.41 (dd, J = 4.0, 13.4 Hz, 1H), 4.28–3.87 (m, 5 H), 3.77 (t, J = 9.2 Hz, 1H), 3.58 (dd, J = 2.6, 9.6 Hz, 1H), 3.41 (t, J = 9.0 Hz, 1H), 3.21 (J = 7.4 Hz, 1H), 2.43–2.31 (m, 4H), 1.66–1.47 (m, 4H), 1.29–1.16 (m, 16H), 0.84–0.78 (m, 6H, CH₃) ppm; ³¹P NMR (81 MHz, D₂O) δ 7.66 (s, 1P) ppm.

1,2-*O***-Dibutyryl***-sn***-glyceryl1**-(**4**-*O***-Phosphoryl**-**D**-*myo***-inositol) Phosphate Sodium Salt (35c).** Preparation as above from protected phosphoinositide **34c** gave final product **35b** (85% yield) as the sodium salt: ¹H NMR (200 MHz, D₂O) δ 5.40–5.26 (m, 1H), 4.37 (dd, J = 3.2, 12.0 Hz, 1H), 4.30–4.17 (m, 2H), 4.12–4.03 (m, 3H), 3.98–3.88 (m, 1H), 3.79 (t, J = 9.6 Hz, 1H), 3.60 (dd, J = 2.6, 9.8 Hz, 1H), 3.43 (t, J = 9.3 Hz, 1H), 2.41–2.31 (m, 4H), 1.69–1.47 (m, 4H), 0.89, 0.87 (2t, J = 7.4 Hz, 6H, CH₃) ppm; ³¹P NMR (81 MHz, D₂O) δ 7.81 (s, 1P), 2.73 (s, 1P) ppm.

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